Technical Manual

PowerPlex® 16 System

INSTRUCTIONS FOR USE OF PRODUCTS DC6530 AND DC6531

This Technical Manual has been revised. Please read it carefully.

- The PowerPlex® 16 System shipped with this Technical Manual incorporates improved manufacturing processes.
- The PowerPlex® 16 Allelic Ladder Mix and PowerPlex® 16 10X Primer Pair Mix included in the kit are not compatible with previous lots of the PowerPlex® 16 System.
- A new matrix must be generated using one of the new matrices: PowerPlex® Matrix Standards 310 (Cat.# DG4640) or PowerPlex® Matrix Standards 3100/3130 (Cat.# DG4650).

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Part# TMD012

PowerPlex® 16 System

All technical literature is available on the Internet at: www.promega.com/tbs/
Please visit the web site to verify that you are using the most current version of this Technical Manual.
Please contact Promega Technical Services if you have questions on use of this system.
E-mail: genetic@promega.com

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I. Description

STR (short tandem repeat) loci consist of short, repetitive sequence elements 3–7 base pairs in length (1–4). These repeats are well distributed throughout the human genome and are a rich source of highly polymorphic markers, which may be detected using the polymerase chain reaction (5–8). Alleles of STR loci are differentiated by the number of copies of the repeat sequence contained within the amplified region and are distinguished from one another using radioactive, silver stain or fluorescence detection following electrophoretic separation.

The PowerPlex® 16 System[a–d] (9,10) allows the co-amplification and three-color detection of sixteen loci (fifteen STR loci and Amelogenin), including Penta E, D18S51, D21S11, TH01, D3S1358, FGA, TPOX, D8S1179, vWA, Amelogenin, Penta D, CSF1PO, D16S539, D7S820, D13S317 and D5S818. One primer for each of the Penta E, D18S51, D21S11, TH01 and D3S1358 loci is labeled with fluorescein (FL); one primer for each of the FGA, TPOX, D8S1179, vWA and Amelogenin loci is labeled with carboxy-tetramethylrhodamine (TMR); and one primer for each of the Penta D, CSF1PO, D16S539, D7S820, D13S317 and D5S818 loci is labeled with 6-carboxy-4´,5´-dichloro-2´,7´-dimethoxy-fluorescein (JOE). All sixteen loci are amplified simultaneously in a single tube and analyzed in a single injection or gel lane.

The PowerPlex® 16 Monoplex System, Penta E (Fluorescein) (Cat.# DC6591) and PowerPlex® 16 Monoplex System, Penta D (JOE) (Cat.# DC6651) are available to amplify the Penta E and Penta D loci, respectively. Each monoplex system allows amplification of a single locus to confirm results obtained with the PowerPlex® 16 System, PowerPlex® 16 BIO System or PowerPlex® 2.1 System. The monoplex systems can also be used to re-amplify DNA samples when one or more of the loci do not amplify initially due to nonoptimal amplification conditions or poor DNA template quality.

The PowerPlex® 16 System is compatible with the ABI PRISM® 310, 3100 and 3100-*Avan*t Genetic Analyzers, Applied Biosystems 3130 and 3130*xl* Genetic Analyzers and ABI PRISM® 377 DNA Sequencer. The protocols presented in this manual were tested at Promega Corporation. Amplification and detection instrumentation may vary. You may need to optimize protocols including cycle number and injection time (or loading volume) for each laboratory instrument. In-house validation should be performed.

The PowerPlex® 16 System provides all of the materials necessary for amplification of STR regions of purified genomic DNA except for AmpliTaq Gold® DNA polymerase. This manual contains separate protocols for use of the PowerPlex® 16 System with the Perkin-Elmer model 480 and GeneAmp® PCR system 9600, 9700 and 2400 thermal cyclers in addition to protocols for separation of amplified products and detection of separated material (Figure 1). Protocols for operation of the fluorescence-detection instruments should be obtained from the instrument manufacturer.

Information on Promega's other fluorescent STR systems and detection of amplified STR fragments using silver staining is available upon request from Promega or online at: **www.promega.com**

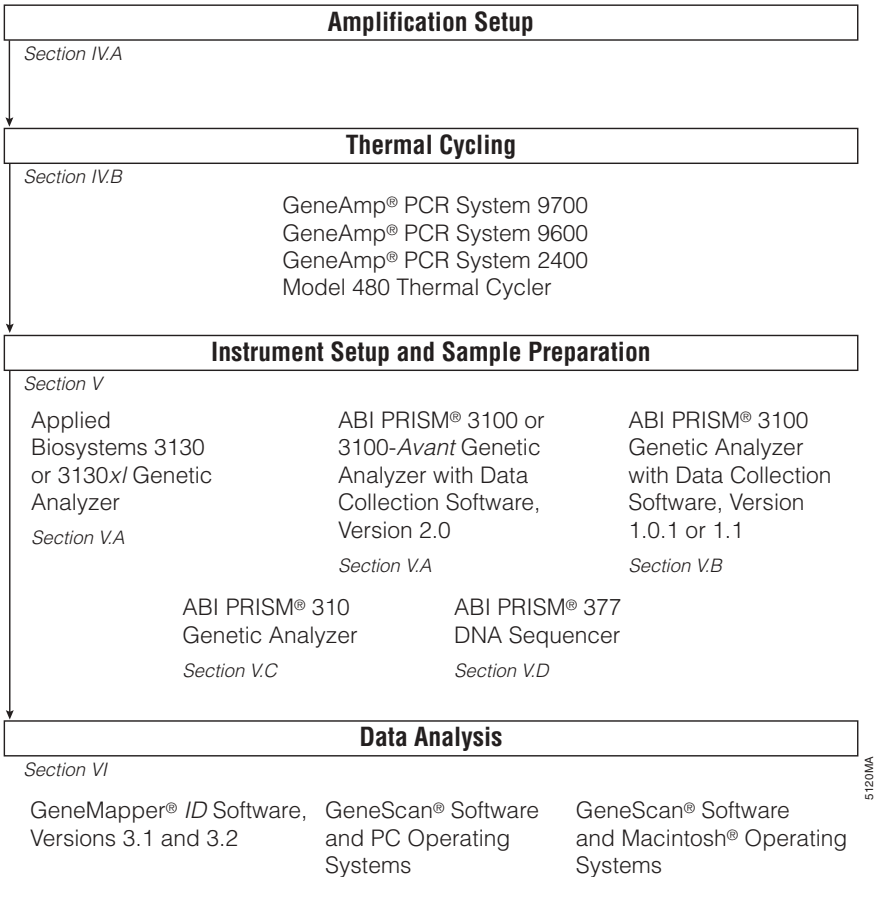

Figure 1. An overview of the PowerPlex® 16 System protocol.

II. Product Components and Storage Conditions

Product	Size	Cat.#
PowerPlex® 16 System	100 reactions	DC6531

Not For Medical Diagnostic Use. Cat.# DC6531 contains sufficient reagents for 100 reactions of 25µl each. Includes:

Pre-amplification Components Box (Blue Label)

1 × 300µl	Gold ST★R 10X Buffer
1 × 250µl	PowerPlex® 16 10X Primer Pair Mix
25µl	9947A DNA (10ng/µl)

Post-amplification Components Box (Beige Label)

1 × 25µl	PowerPlex® 16 Allelic Ladder Mix
1× 150µl	Internal Lane Standard (ILS) 600
1	Protocol

Product	Size	Cat.#
PowerPlex® 16 System	400 reactions	DC6530

Not For Medical Diagnostic Use. Cat.# DC6530 contains sufficient reagents for 400 reactions of 25µl each. Includes:

Pre-amplification Components Box (Blue Label)

4 × 300µl	Gold ST★R 10X Buffer
4 × 250µl	PowerPlex® 16 10X Primer Pair Mix
25µl	9947A DNA (10ng/µl)

Post-amplification Components Box (Beige Label)

4 × 25µl	PowerPlex® 16 Allelic Ladder Mix
4 × 150µl	Internal Lane Standard (ILS) 600
1	Protocol

The PowerPlex® 16 Allelic Ladder Mix is provided in a separate, sealed bag for shipping. This component should be moved to the post-amplification box after opening.

Storage Conditions: Store all components at –20°C in a nonfrost-free freezer. The PowerPlex® 16 10X Primer Pair Mix, PowerPlex® 16 Allelic Ladder Mix and the Internal Lane Standard 600 are light-sensitive and must be stored in the dark. We strongly recommend that pre-amplification and post-amplification reagents be stored and used separately with different pipettes, tube racks, etc.

Available Separately

Product	Size	Cat.#
Blue Dextran Loading Solution*	3ml	DV4351
PowerTyper™ Macros (Release 2.0)**	1 CD-ROM	DG3470

*For Laboratory Use.
**Not For Medical Diagnostic Use.

The PowerTyper™ Macros (Release 2.0), for use with Genotyper® software, are available from Promega. This CD-ROM contains the file "PowerTyper™ 16 Macro (Release 2.0)" for use with the PowerPlex® 16 System. The macros can also be downloaded at: **www.promega.com/geneticidtools/**

The proper panel and bin files for use with GeneMapper® *ID* software can be obtained from the Promega web site at: **www.promega.com/geneticidtools/panels_bins/**

Matrix standards are required for initial setup of the color separation matrix. The matrix standards are sold separately and are available for the ABI PRISM® 310 Genetic Analyzer and 377 DNA Sequencer (PowerPlex® Matrix Standards, 310) and the ABI PRISM® 3100 and 3100-*Avant* Genetic Analyzers and Applied Biosystems 3130 and 3130*xl* Genetic Analyzers (PowerPlex® Matrix Standards, 3100/3130). See Section IX.I for ordering information.

III. Before You Begin

III.A. Precautions

The application of PCR-based typing for forensic or paternity casework requires validation studies and quality-control measures that are not contained in this manual (11,12). The quality of the purified DNA sample, as well as small changes in buffers, ionic strength, primer concentrations, choice of thermal cycler and thermal cycling conditions, can affect the success of a PCR amplification. We suggest strict adherence to recommended procedures for amplification, as well as electrophoresis and fluorescence detection.

PCR-based STR analysis is subject to contamination by very small amounts of nontemplate human DNA. Extreme care should be taken to avoid cross-contamination when preparing sample DNA, handling primer pairs, assembling amplification reactions and analyzing amplification products. Reagents and materials used prior to amplification (Gold ST★R 10X Buffer and PowerPlex® 16 10X Primer Pair Mix) are provided in a separate box and should be stored separately from those used following amplification (PowerPlex® 16 Allelic Ladder Mix and Internal Lane Standard 600). Always include a negative control reaction (i.e., no template) to detect reagent contamination. We highly recommend the use of gloves and aerosol-resistant pipette tips (e.g., ART® tips, Section IX.I).

Some of the reagents used in the analysis of STR products are potentially hazardous and should be handled accordingly. Table 1 describes the potential hazards associated with such reagents.

Table 1. Hazardous Reagents.

Reagents for ABI PRISM® 310, 3100 and 3100-*Avant* Genetic Analyzers and Applied Biosystems 3130 and 3130*xl* Genetic Analyzers	Hazard
formamide	irritant, teratogen

Reagents for ABI PRISM® 377 DNA Sequencer	Hazard
acrylamide (Long Ranger® gel solution)	suspected carcinogen, toxic
ammonium persulfate	oxidizer, corrosive
formamide (contained in the Blue Dextran Loading Solution)	irritant, teratogen
TEMED	corrosive, flammable
urea	irritant

III.B. Matrix Standardization or Spectral Calibration

Proper generation of a matrix file is critical to evaluate multicolor systems with the ABI PRISM® 310, 3100 and 3100-*Avant* Genetic Analyzers and Applied Biosystems 3130 and 3130*xl* Genetic Analyzers. A matrix must be generated for each individual instrument.

The PowerPlex® Matrix Standards, 310 (Cat.# DG4640), is required for matrix standardization for the ABI PRISM® 310 Genetic Analyzer and ABI PRISM® 377 DNA Sequencer. For best results, the PowerPlex® Matrix Standards, 3100/3130 (Cat.# DG4650), should not be used to generate a matrix on the ABI PRISM® 310 Genetic Analyzer.

The PowerPlex® Matrix Standards, 3100/3130 (Cat.# DG4650), is required for spectral calibration on the ABI PRISM® 3100 and 3100-*Avant* Genetic Analyzers and Applied Biosystems 3130 and 3130*xl* Genetic Analyzers. The PowerPlex® Matrix Standards, 310 (Cat.# DG4640), cannot be used to generate a matrix on these instruments.

For protocols and additional information on matrix standardization, see the *PowerPlex® Matrix Standards, 310, Technical Bulletin* #TBD021, which is supplied with Cat.# DG4640. For protocols and additional information about spectral calibration, see the *PowerPlex® Matrix Standards, 3100/3130, Technical Bulletin* #TBD022, which is supplied with Cat.# DG4650. These manuals are available upon request from Promega or online at: **www.promega.com/tbs/**

IV. Protocols for DNA Amplification Using the PowerPlex® 16 System

Materials to Be Supplied by the User

- thermal cycler model 480 or GeneAmp® PCR System 9600, 9700 or 2400 (Applied Biosystems)
- microcentrifuge
- 0.5ml or 0.2ml thin-walled microcentrifuge tubes or MicroAmp® optical 96-well reaction plate (Applied Biosystems)
- 1.5ml amber-colored microcentrifuge tubes (Fisher Cat.# 05-402-26)
- aerosol-resistant pipette tips (see Section IX.I)
- AmpliTaq Gold® DNA polymerase (Applied Biosystems)
- Nuclease-Free Water (Cat.# P1193)
- Mineral Oil (Cat.# DY1151, for use with the model 480 thermal cycler)

We routinely amplify 0.5–1ng of template DNA in a 25µl reaction volume using the protocols detailed below. Preferential amplification of the smaller loci can occur. Expect to see high peak heights in the smaller loci and relatively lower peak heights in the larger loci if more than the recommended amount of template is used. Reduce the amount of template DNA or the number of cycles to correct this.

The PowerPlex® 16 System is optimized for the GeneAmp® PCR System 9700 thermal cycler. Amplification protocols for the GeneAmp® PCR Systems 9600 and 2400 thermal cyclers and the Perkin-Elmer model 480 thermal cycler are provided.

IV.A. Amplification Setup

The use of gloves and aerosol-resistant pipette tips is highly recommended to prevent cross-contamination. Keep all pre-amplification and post-amplification reagents in separate rooms. Prepare amplification reactions in a room dedicated for reaction setup. Use equipment and supplies dedicated for amplification setup.

Meticulous care must be taken to ensure successful amplification. A guide to amplification troubleshooting is provided in Section VII.A.

1. Thaw the Gold ST★R 10X Buffer and PowerPlex® 16 10X Primer Pair Mix.

   Notes:

   1. Mix reagents by vortexing for 15 seconds before each use. Do not centrifuge the 10X Primer Pair Mix, as this may cause the primers to be concentrated at the bottom of the tube.
   2. A precipitate may form in the Gold ST★R 10X Buffer. If this occurs, warm the solution briefly at 37°C, then vortex until the precipitate is in solution.

2. Determine the number of reactions to be set up. This should include positive and negative control reactions. Add 1 or 2 reactions to this number to compensate for pipetting error. While this approach does waste a small amount of each reagent, it ensures that you will have enough PCR master mix for all samples. It also ensures that each reaction contains the same master mix.

3. Place one clean, 0.2ml or 0.5ml reaction tube for each reaction into a rack, and label appropriately. Alternatively, use a MicroAmp® plate, and label appropriately.

   Note: If using the GeneAmp® PCR System 9600, 9700 or 2400 thermal cyclers, use 0.2ml MicroAmp® 8-strip reaction tubes or MicroAmp® plate. For the Perkin-Elmer model 480 thermal cycler, we recommend the 0.5ml GeneAmp® thin-walled reaction tubes.

4. Add the final volume of each reagent listed in Table 2 into a sterile, 1.5ml amber-colored tube. Mix gently.

   Table 2 shows the component volumes per reaction. A worksheet to calculate the required amount of each component of the PCR master mix is provided in Section IX.F (Table 10).

   Table 2. PCR Master Mix for the PowerPlex® 16 System.

PCR Master Mix Component[1]	Volume Per Reaction
nuclease-free water	to a final volume of 25.0µl
Gold ST★R 10X Buffer	2.5µl
PowerPlex® 16 10X Primer Pair Mix	2.5µl
AmpliTaq Gold® DNA polymerase[2]	0.8µl (4u)
template DNA (0.5–1ng)[3]	up to 19.2µl
total reaction volume	**25µl**

   [1]Add nuclease-free water to the PCR master mix first, then add Gold ST★R 10X Buffer, PowerPlex® 16 10X Primer Pair Mix and AmpliTaq Gold® DNA polymerase. The template DNA will be added at Step 6.

   [2]Assumes the AmpliTaq Gold® DNA polymerase is at 5u/µl. If the enzyme concentration is different, the volume of enzyme must be adjusted accordingly.

   [3]Store DNA templates in nuclease-free water or TE[-4] buffer (10mM Tris HCl [pH 8.0], 0.1mM EDTA). If the DNA template is stored in TE buffer that is not pH 8.0 or contains a higher EDTA concentration, the volume of the DNA sample added should not exceed 20% of the final reaction volume. PCR amplification efficiency and quality can be greatly altered by changes in pH (due to added Tris-HCl), available magnesium concentration (due to chelation by EDTA) or other PCR inhibitors, which may be present at low concentrations depending on the source of the template DNA and the extraction procedure used.

   ! Amplification of >1ng of DNA template results in an imbalance in peak heights from locus to locus. The smaller loci show greater amplification yield than the larger loci. Reducing the number of cycles in the amplification program by 2 to 4 cycles (i.e., 10/20 or 10/18 cycling) can improve locus-to-locus balance.

5. Pipet PCR master mix into each reaction tube.

6. Pipet the template DNA (0.5–1ng) for each sample into the respective tube containing PCR master mix.

7. For the positive amplification control, dilute the 9947A DNA positive control to 0.5ng in the desired template DNA volume. Pipet 0.5ng of the diluted DNA into a reaction tube containing the PCR master mix.

8. For the negative amplification control, pipet nuclease-free water (instead of template DNA) into a reaction tube containing the PCR master mix.

9. If using the GeneAmp® PCR System 9600, 9700 or 2400 thermal cycler and MicroAmp® reaction tubes or plates, no addition of mineral oil to the reaction tubes is required. However, if using the model 480 thermal cycler and GeneAmp® reaction tubes, add one drop of mineral oil to each tube before closing.

   Note: Allow the mineral oil to flow down the side of the tube and form an overlay to limit sample loss or cross-contamination due to splattering.

IV.B. Amplification Thermal Cycling

This manual contains protocols for use of the PowerPlex® 16 System with the Perkin-Elmer model 480 and GeneAmp® PCR system 9600, 9700 and 2400 thermal cyclers. For information about other thermal cyclers, please contact Promega Technical Services by e-mail: **genetic@promega.com**

Amplification and detection instrumentation may vary. You may need to optimize protocols including cycle number and injection time (or loading volume) for each laboratory instrument. Testing at Promega Corporation shows that 10/22 cycles work well for 0.5–1ng of purified DNA templates. For higher amounts of input DNA (i.e., FTA® paper) or to decrease sensitivity, fewer cycles, such as 10/16, 10/18 or 10/20, should be evaluated. In-house validation should be performed.

1. Place the tubes or MicroAmp® plate in the thermal cycler.

2. Select and run a recommended protocol. The preferred protocols for use with the GeneAmp® PCR System 9600, 9700 and 2400 thermal cyclers and the Perkin-Elmer model 480 thermal cycler are provided below.

3. After completion of the thermal cycling protocol, store the samples at –20°C in a light-protected box.

   Note: Storage of amplified samples at 4°C or higher may produce degradation products.

Protocol for the GeneAmp® PCR System 9700 Thermal Cycler[1]	Protocol for the GeneAmp® PCR System 2400 Thermal Cycler
95°C for 11 minutes, then:	95°C for 11 minutes, then:
96°C for 1 minute, then:	96°C for 1 minute, then:
ramp 100% to 94°C for 30 seconds ramp 29% to 60°C for 30 seconds ramp 23% to 70°C for 45 seconds for 10 cycles, then:	ramp 100% to 94°C for 30 seconds ramp 100% to 60°C for 30 seconds ramp 23% to 70°C for 45 seconds for 10 cycles, then:
ramp 100% to 90°C for 30 seconds ramp 29% to 60°C for 30 seconds ramp 23% to 70°C for 45 seconds for 22 cycles, then:	ramp 100% to 90°C for 30 seconds ramp 100% to 60°C for 30 seconds ramp 23% to 70°C for 45 seconds for 22 cycles, then:
60°C for 30 minutes	60°C for 30 minutes
4°C soak	4°C soak

Protocol for the GeneAmp® PCR System 9600 Thermal Cycler	Protocol for the Perkin-Elmer Model 480 Thermal Cycler
95°C for 11 minutes, then:	95°C for 11 minutes, then:
96°C for 1 minute, then:	96°C for 2 minutes, then:
94°C for 30 seconds ramp 68 seconds to 60°C (hold for 30 seconds) ramp 50 seconds to 70°C (hold for 45 seconds) for 10 cycles, then:	94°C for 1 minute 60°C for 1 minute 70°C for 1.5 minutes for 10 cycles, then:
90°C for 30 seconds ramp 60 seconds to 60°C (hold for 30 seconds) ramp 50 seconds to 70°C (hold for 45 seconds) for 22 cycles, then:	90°C for 1 minute 60°C for 1 minute 70°C for 1.5 minutes for 22 cycles, then:
60°C for 30 minutes	60°C for 30 minutes
4°C soak	4°C soak

[1]When running the GeneAmp® PCR System 9700 thermal cycler, use the ramp rate for the GeneAmp® PCR System 9600 thermal cycler.

V. Instrument Setup and Sample Preparation

V.A. Detection of Amplified Fragments Using the ABI PRISM® 3100 or 3100-*Avant* Genetic Analyzer with Data Collection Software, Version 2.0, and the Applied Biosystems 3130 or 3130*xl* Genetic Analyzer

Materials to Be Supplied by the User

- 95°C dry heating block, water bath or thermal cycler
- crushed ice or ice-water bath
- aerosol-resistant pipette tips
- 3100 or 3130 capillary array, 36cm
- performance optimized polymer 4 (POP-4™) for the 3100 or 3130
- 10X genetic analyzer buffer with EDTA
- MicroAmp® optical 96-well plate and septa
- Hi-Di™ formamide (Applied Biosystems Cat.# 4311320)
- PowerPlex® Matrix Standards, 3100/3130 (Cat.# DG4650)

The quality of the formamide is critical. Use Hi-Di™ formamide with a conductivity less than 100µS/cm. Freeze formamide in aliquots at –20°C. Multiple freeze-thaw cycles or long-term storage at 4°C may cause a breakdown of the formamide. Formamide with a conductivity greater than 100µS/cm may contain ions that compete with DNA during injection. This results in lower peak heights and reduced sensitivity. A longer injection time may not increase the signal.

Caution: Formamide is an irritant and a teratogen; avoid inhalation and contact with skin. Read the warning label, and take appropriate precautions when handling this substance. Always wear gloves and safety glasses when working with formamide.

Sample Preparation

1. Prepare a loading cocktail by combining and mixing the internal lane standard and Hi-Di™ formamide as follows:

   [(0.5µl ILS 600) × (# injections)] + [(9.5µl Hi-Di™ formamide) × (# injections)]

   Note: The volume of internal lane standard used in the loading cocktail can also be increased or decreased to adjust the intensity of the size standard peaks. The optimal peak height for the 100-base fragment of the internal lane standard is 500–1,000RFU. If the peak heights are too low, we recommend altering the formamide/internal lane standard mix to contain 1.0µl of ILS 600 and 9.0µl of Hi-Di™ formamide. If the peak heights are too high, we recommend altering the loading cocktail to contain 0.25µl of ILS 600 and 9.75µl of formamide.

2. Mix for 10–15 seconds using a vortex mixer.

3. Pipet 10µl of formamide/internal lane standard mix into each well.

V.A. Detection of Amplified Fragments Using the ABI PRISM® 3100 or 3100-*Avant* Genetic Analyzer with Data Collection Software, Version 2.0, and the Applied Biosystems 3130 or 3130*xl* Genetic Analyzer (continued)

4. Add 1µl of amplified sample (or 1µl of the allelic ladder mix). Cover wells with appropriate septa.

   Note: Instrument detection limits vary; therefore, injection time or the amount of product mixed with loading cocktail may need to be increased or decreased. Use the "Module Manager" in the data collection software to modify the injection time or voltage in the run module. If the peak heights are higher than desired, the samples can be diluted in Gold ST★R 1X Buffer before mixing with loading cocktail. This may result in uneven allele peak heights across loci. For best results, use less DNA template in the amplification reactions or reduce the number of cycles in the amplification program by 2–4 cycles to achieve the desired signal intensity.

5. Centrifuge plate briefly to remove air bubbles from the wells if necessary.

6. Denature samples at 95°C for 3 minutes, then immediately chill on crushed ice or in an ice-water bath for 3 minutes. Denature the samples just prior to loading the instrument.

Instrument Preparation

Refer to the instrument users' manual for instructions on cleaning, installing the capillary array, performing a spatial calibration and adding polymer.

Analyze the samples as described in the user's manual for the ABI PRISM® 3100 or 3100-*Avant* Genetic Analyzer with data collection software, version 2.0, and the Applied Biosystems 3130 or 3130*xl* Genetic Analyzer with the following exceptions.

1. In the Module Manager, select "New". Select "Regular" in the Type drop-down list, and select "HIDFragmentAnalysis36_POP4" in the Template drop-down list. Confirm that the injection time is 5 seconds and the injection voltage is 3kV. Lengthen the run time to 2,000 seconds. Give a descriptive name to your run module, and select "OK".

   Note: Sensitivities of instruments may vary. The injection time and voltage may be adjusted in the Module Manager. A suggested range for the injection time is 3–22 seconds and for the injection voltage is 1–3kV.

2. In the Protocol Manager, select "New". Type a name for your protocol. Select "Regular" in the Type drop-down list, and select the run module you created in the previous step in the Run Module drop-down list. Lastly, select "F" in the Dye-Set drop-down list. Select "OK.

3. In the Plate Manager, create a new plate record as described in the instrument user's manual. In the dialog box that appears, select "GeneMapper—Generic" in the Application drop-down list, and select the appropriate plate type (96-well). Add entries in the owner and operator windows, and select "OK".

   Note: If autoanalysis of sample data is desired, refer to the instrument user's manual for instructions.

4. In the GeneMapper® plate record, enter sample names in the appropriate cells. Scroll to the right. In the "Results group 1" column, select the desired results group. In the "Instrument Protocol 1" column, select the protocol you created in Step 2. Be sure this information is present for each row that contains a sample name. Select "OK".

   Note: To create a new results group, select "New" in the drop-down menu in the results group column. Select the General tab, and enter a name. Select the Analysis tab, and select "GeneMapper—Generic" in the Analysis type drop-down list.

5. Place samples in the instrument, and close the instrument doors.

6. In the spectral viewer, confirm that dye set F is active, and set the correct active calibration for dye set F.

7. In the run scheduler, locate the plate record that you just created in Steps 3 and 4, and click once on the name to highlight it.

8. Once the plate record is highlighted, click the plate graphic that corresponds to the plate on the autosampler that contains your amplified samples.

9. When the plate record is linked to the plate, the plate graphic will change from yellow to green, and the green Run Instrument arrow becomes enabled.

10. Click on the green Run Instrument arrow on the toolbar to start the sample run.

11. Monitor electrophoresis by observing the run, view, array or capillaries viewer window in the data collection software. Each injection will take approximately 45 minutes.

V.B. Detection of Amplified Fragments Using the ABI PRISM® 3100 Genetic Analyzer with Data Collection Software, Version 1.0.1 or 1.1

Materials to Be Supplied by the User

- 95°C dry heating block, water bath or thermal cycler
- crushed ice or ice-water bath
- aerosol-resistant pipette tips
- 3100 capillary array, 36cm
- performance optimized polymer 4 (POP-4™) for the 3100
- 10X genetic analyzer buffer with EDTA
- MicroAmp® optical 96-well plate and septa for the 3100
- Hi-Di™ formamide (Applied Biosystems Cat.# 4311320)
- PowerPlex® Matrix Standards, 3100/3130 (Cat.# DG4650)

The quality of the formamide is critical. Use Hi-Di™ formamide with a conductivity less than 100µS/cm. Freeze formamide in aliquots at –20°C. Multiple freeze-thaw cycles or long-term storage at 4°C may cause a breakdown of the formamide. Formamide with a conductivity greater than 100µS/cm may contain ions that compete with DNA during injection. This results in lower peak heights and reduced sensitivity. A longer injection time may not increase the signal.

Caution: Formamide is an irritant and a teratogen; avoid inhalation and contact with skin. Read the warning label, and take appropriate precautions when handling this substance. Always wear gloves and safety glasses when working with formamide.

Sample Preparation

1. Prepare a loading cocktail by combining and mixing the internal lane standard and Hi-Di™ formamide as follows:

   [(0.5µl ILS 600) × (# injections)] + [(9.5µl Hi-Di™ formamide) × (# injections)]

   Note: The volume of internal lane standard used in the loading cocktail can also be increased or decreased to adjust the intensity of the size standard peaks. The optimal peak height for the 100-base fragment of the internal lane standard is 500–1,000RFU. If the peak heights are too low, we recommend altering the formamide/internal lane standard mix to contain 1.0µl of ILS 600 and 9.0µl of Hi-Di™ formamide. If the peak heights are too high, we recommend altering the loading cocktail to contain 0.25µl of ILS 600 and 9.75µl of formamide.

2. Mix for 10–15 seconds using a vortex mixer.

3. Pipet 10µl of formamide/internal lane standard mix into each well.

4. Add 1µl of amplified sample (or 1µl of the allelic ladder mix). Cover wells with appropriate septa.

   Note: Instrument detection limits vary; therefore, injection time or the amount of product mixed with loading cocktail may need to be increased or decreased. Use the Module Editor under "Tools" to modify the injection time in the run module. If the peak heights are higher than desired, the samples can be diluted in Gold ST★R 1X Buffer before mixing with loading cocktail. The use of too much template DNA may result in uneven allele peak heights across loci. For best results, use less DNA template in the amplification reactions or reduce the number of cycles in the amplification program by 2–4 cycles to achieve the desired signal intensity.

5. Centrifuge plate briefly to remove air bubbles from the wells if necessary.

6. Denature samples at 95°C for 3 minutes, then immediately chill on crushed ice or in an ice-water bath for 3 minutes. Denature the samples just prior to loading the instrument.

Instrument Preparation

Refer to the *ABI PRISM® 3100 Genetic Analyzer User's Manual* for instructions on cleaning the blocks, installing the capillary array, performing a spatial calibration and adding polymer to the reserve syringe.

1. Open the ABI PRISM® 3100 data collection software.

2. Change the "GeneScan36_POP4DefaultModule" module run time to 2,000 seconds.

3. Change the injection voltage to 3kV.

4. Change the injection time to 11 seconds.

   Note: Sensitivities of instruments may vary. The injection time and voltage may be adjusted in the Module Manager. A suggested range for the injection time is 3–22 seconds and for the injection voltage is 1–3kV.

5. Save the module with a new name (e.g., GeneScan36_POP4PowerPlex16_3kV_11secs_2000). Use this as the initial run module for all runs.

6. Open a new plate record. Name the plate, and select "GeneScan". Select the plate size (96-well). Select "Finish".

7. Complete the plate record spreadsheet for the wells you have loaded. Enter appropriate information into the "sample name" and "color info" columns. For allelic ladder samples, insert the word "ladder" into the "color info" column for the blue, yellow and green dye colors. This information must be entered to successfully analyze data with the PowerTyper™ 16 Macro (Release 2.0).

8. In the "BioLIMS Project" column, select "3100_Project1" from the pull-down menu.

9. In the "Dye Set" column, select "Z" from the pull-down menu.

10. When using the ABI PRISM® 3100 data collection software version 1.0.1 or 1.1, select "GeneScan36_POP4PowerPlex16_3kV_11secs_2000" from the pull-down menu in the "Run Module 1" column.

11. To collect the data without autoanalyzing, select "No Selection" in the "Analysis Module 1" column. Analysis parameters can be applied after data collection and during data analysis using the GeneScan® analysis software.

12. Select "OK". This new plate record will appear in the pending plate records table on the plate setup page of the collection software.

13. Place samples in the instrument, and close the instrument doors.

14. Locate the pending plate record that you just created, and click once on the name.

15. Once the pending plate record is highlighted, click on the plate graphic that corresponds to the plate on the autosampler that contains your amplified samples to link the plate to the plate record.

16. When the plate record is linked to the plate, the plate graphic will change from yellow to green, the plate record moves from the pending plate records table to the linked plate records table, and the Run Instrument button becomes enabled.

17. Select "Run Instrument" on the toolbar to start the sample run.

18. Monitor electrophoresis by observing the run, status, array and capillary views windows in the collection software. Each injection will take approximately 45 minutes.

V.C. Detection of Amplified Fragments Using the ABI PRISM® 310 Genetic Analyzer

Materials to Be Supplied by the User

- 95°C dry heating block, water bath or thermal cycler
- 310 capillaries, 47cm × 50µm
- performance optimized polymer 4 (POP-4™)
- glass syringe (1ml)
- 10X genetic analyzer buffer with EDTA
- sample tubes and septa
- aerosol-resistant pipette tips
- Hi-Di™ formamide (Applied Biosystems Cat.# 4311320)
- PowerPlex® Matrix Standards, 310 (Cat.# DG4640)
- crushed ice or ice-water bath

The quality of the formamide is critical. Use Hi-Di™ formamide with a conductivity less than 100µS/cm. Freeze formamide in aliquots at –20°C. Multiple freeze-thaw cycles or long-term storage at 4°C may cause a breakdown of the formamide. Formamide with a conductivity greater than 100µS/cm may contain ions that compete with DNA during injection. This results in lower peak heights and reduced sensitivity. A longer injection time may not increase the signal.

Caution: Formamide is an irritant and a teratogen; avoid inhalation and contact with skin. Read the warning label, and take appropriate precautions when handling this substance. Always wear gloves and safety glasses when working with formamide.

Sample Preparation

1. Prepare a loading cocktail by combining the Internal Lane Standard 600 (ILS 600) and Hi-Di™ formamide as follows:

   [(1.0µl ILS 600) × (# injections)] + [(24.0µl Hi-Di™ formamide) × (# injections)]

   Note: The volume of internal lane standard used in the loading cocktail can also be increased or decreased to adjust the intensity of the size standard peaks. The optimal peak height for the 100-base fragment of the internal lane standard is 500–1,000RFU. If the peak heights are too high, we recommend altering the loading cocktail to contain 0.5µl of ILS 600 and 24.5µl of Hi-Di™ formamide.

2. Mix for 10–15 seconds using a vortex mixer.

3. Combine 25.0µl of the prepared loading cocktail and 1.0µl of amplified sample.

   Note: Instrument detection limits vary; therefore, injection time or the amount of product mixed with loading cocktail may need to be increased or decreased. If the peak heights are higher than desired, the samples can be diluted in Gold ST★R 1X Buffer before mixing with loading cocktail. This may result in uneven allele peak heights across loci. For best results, use less template DNA in the amplification reactions or reduce the number of cycles in the amplification program by 2–4 cycles (i.e., 10/18 or 10/20 cycling).

4. Combine 25.0µl of the prepared loading cocktail and 1.0µl of the PowerPlex® 16 Allelic Ladder Mix.
5. Centrifuge the sample tubes briefly to remove air bubbles from the wells if necessary.
6. Denature the samples and ladder by heating at 95°C for 3 minutes, and immediately chill on crushed ice or in an ice-water bath for 3 minutes. Denature the samples just prior to loading.
7. Assemble the tubes in the appropriate autosampler tray (48- or 96-tube).
8. Place the autosampler tray in the instrument, and close the instrument doors.

Instrument Preparation

Refer to the instrument users' manual for instructions on cleaning the pump block, installing the capillary, calibrating the autosampler and adding polymer to the syringe.

1. Open the ABI PRISM® 310 data collection software.
2. Prepare a GeneScan® sample sheet as described in the *ABI PRISM® 310 Genetic Analyzer User's Manual*. Enter the appropriate sample information in the "sample info" column.

   For rows containing the PowerPlex® 16 Allelic Ladder Mix, insert the word "ladder" in the "sample info" column for the blue dye color, yellow dye color and green dye color. This information must be entered to successfully analyze your data using the PowerTyper™ 16 Macro (Release 2.0).
3. Create a new GeneScan® injection list. Select the appropriate sample sheet by using the pull-down menu.
4. Select the "GS STR POP4 (1ml) A" Module using the pull-down menu. Change the injection time to 3 seconds and the run time to 30 minutes. Keep the settings for the remaining parameters as shown below:

Inj. Secs:	3
Inj. kV:	15.0
Run kV:	15.0
Run °C:	60
Run Time:	30

You may need to optimize the injection time for individual instruments. Injection times of 2–5 seconds are suggested for samples that contain 1ng of template DNA.

Note: Migration of fragments may vary slightly over the course of a long ABI PRISM® 310 Genetic Analyzer run. This may be due to changes in temperature or changes in the column. When analyzing many samples, injections of the allelic ladder at different times throughout the run can aid in accurately genotyping the samples.

V.C. Detection of Amplified Fragments Using the ABI PRISM® 310 Genetic Analyzer (continued)

5. Select the appropriate matrix file (Section III.B).
6. To analyze the data automatically, select the auto analyze checkbox and the appropriate analysis parameters and size standard. Refer to the *ABI PRISM® 310 Genetic Analyzer User's Manual* for specific information on these options.
7. After loading the sample tray and closing the doors, select "Run" to start the capillary electrophoresis system.
8. Monitor the electrophoresis by observing the raw data and status windows. Each sample will take approximately 40 minutes for syringe pumping, sample injection and sample electrophoresis.

V.D. Detection of Amplified Fragments Using the ABI PRISM® 377 DNA Sequencer

Materials to Be Supplied by the User

(Solution compositions are provided in Section IX.H.)

- Long Ranger® gel solution (Cambrex Cat.# 50611) or Long Ranger Singel® pack for ABI 377-36cm (Cambrex Cat.# 50691)
- 10% Ammonium Persulfate (Cat.# V3131)
- TEMED
- Urea (Cat.# V3171)
- TBE 10X buffer
- Nalgene® tissue culture filter (0.2 micron)
- 36cm front and rear glass plates
- 36cm gel spacers (0.2mm thick)
- 36-well sharkstooth comb or 34-well square-tooth comb (0.2mm thick)
- clamps (e.g., large office binder clamps)
- gel-loading pipette tips
- aerosol-resistant pipette tips (Section IX.I.)
- Liqui-Nox® or other detergent
- PowerPlex® Matrix Standards, 310 (Cat.# DG4640)
- Blue Dextran Loading Solution (Cat.# DV4351)
- crushed ice or ice-water bath
- 95°C dry heating block, water bath or thermal cycler

Caution: Acrylamide (Long Ranger® gel solution) is a neurotoxin and suspected carcinogen; avoid inhalation and contact with skin. Read the warning label, and take appropriate precautions when handling this substance. Always wear gloves and safety glasses when working with acrylamide solutions.

Polyacrylamide Gel Preparation

The following protocol is for preparation of a 36cm denaturing polyacrylamide gel for use with the ABI PRISM® 377 DNA sequencer. Low-fluorescence glass plates are recommended and may be obtained from the instrument manufacturer.

1. Thoroughly clean the glass plates with hot water and a 1% Liqui-Nox® solution. Rinse extremely well using deionized water. Allow the glass plates to air-dry in a dust-free environment.
2. Assemble the glass plates by placing 0.2mm side gel spacers between the front and rear glass plates. Hold the plates together using binder clamps (4 clamps on each side). Place the assembly horizontally on a test tube rack or similar support.
3. Prepare a 5% Long Ranger® acrylamide gel (total of 50ml) by combining the ingredients listed in Table 3. Stir the solution until the urea has dissolved.

Table 3. Preparation of a 5% Long Ranger® Polyacrylamide Gel.

Component	5% Gel	Final Concentration
urea	18g	6M
deionized water	26ml	–
10X TBE	5ml	1X
50% Long Ranger® gel solution	5ml	5%
total volume	50ml	

Note: Long Ranger Singel® Packs may be used.

4. Filter the acrylamide solution through a 0.2 micron filter (e.g., Nalgene® tissue culture filter), and degas for 5 minutes.
5. Add 35µl of TEMED and 250µl of fresh 10% ammonium persulfate to 50ml of acrylamide solution, and mix gently.
6. Using a disposable 60cc syringe, pour the gel by starting at the well end of the plates and carefully injecting the acrylamide between the horizontal glass plates. Allow the solution to fill the top width of the plates. While maintaining a constant flow of solution, gently tap the glass plates to assist the movement of solution to the bottom of the plates and to prevent the formation of bubbles.
7. Insert a 36-well sharkstooth comb or 34-well square-tooth comb between the glass plates. Sharkstooth combs with 64 or 96 wells may also be used.
8. Secure the comb with 3 evenly spaced clamps.
9. Keep the remaining acrylamide solution as a polymerization control.
10. Allow polymerization to proceed for at least 2 hours. Check the polymerization control to be sure that polymerization has occurred.

    Note: The gel may be stored overnight if a paper towel saturated with deionized water and plastic wrap are placed around the top and bottom to prevent the gel from drying out (crystallization of the urea will destroy the gel).

Instrument Preparation

1. Open the ABI PRISM® 377 data collection software.
2. Prepare a sample sheet as described in the *GeneScan® Analysis Software User's Manual*. Enter the appropriate sample information in the "sample info" column.

   For lanes containing the PowerPlex® 16 Allelic Ladder Mix, insert the word "ladder" in the "sample info" column for the blue dye color, yellow dye color and green dye color. This information must be entered to successfully analyze your data using the PowerTyper™ 16 Macro (Release 2.0).
3. Create a new GeneScan® run, and use the following settings:

Plate Check Module:	Plate Check A
PreRun Module:	PR GS 36A-2400
Run Module:	GS 36A-2400
Collect time:	3 hours
Well-to-Read distance:	36cm
4. Select the appropriate sample sheet and comb selection by using the pull-down menus.
5. Select the appropriate gel matrix file (Section III.B).

Gel Pre-run

1. Remove the clamps from the polymerized acrylamide gel. If necessary, clean any excess acrylamide from the glass plates with paper towels saturated with deionized water.
2. Shave any excess polyacrylamide away from the comb, and remove the comb. If using a sharkstooth comb, carefully insert the sharkstooth comb teeth into the gel approximately 1–2mm.
3. Position the gel/glass plate unit in the 377 cassette.
4. Secure the cassette in the instrument, and perform a plate check as recommended in the *ABI PRISM® 377 DNA Sequencer User's Manual*. If the horizontal line graph is not flat, remove the cassette, clean the plate surface and repeat the plate check.
5. Add TBE 1X buffer to the top and bottom buffer chambers of the instrument.
6. Using a 60cc syringe filled with buffer, remove any air bubbles from the well area of the gel, and place the lid on the upper buffer chamber. Using a syringe fitted with a bent 18-gauge needle, remove any air bubbles from the bottom of the gel.
7. Attach the heating plate, connect the water tubing, attach all electrodes, close the instrument door and select "PreRun". Allow the gel to pre-run for 15–20 minutes or until the gel temperature is at least 40°C. Open the status window to monitor the temperature of the gel.
8. Prepare the sample and allelic ladder samples during the gel pre-run.

Sample Preparation and Loading

1. Prepare a loading cocktail by combining and mixing the ILS 600 and Blue Dextran Loading Solution as follows:

   [(0.5µl ILS 600) × (# lanes)] + [(1.5µl Blue Dextran Loading Solution) × (# lanes)]

   Note: The volume of internal lane standard used in the loading cocktail can also be increased or decreased to adjust the intensity of the size standard peaks.

2. Vortex for 10–15 seconds.

3. Combine 2.0µl of the prepared loading cocktail and 1.0µl of amplified sample.

   Note: Instrument detection limits vary; therefore, the amount of product mixed with loading cocktail may need to be increased or decreased. If the peak heights are higher than desired, the samples can be diluted in Gold ST★R 1X Buffer before mixing with loading cocktail. This may result in uneven allele peak heights across loci. For best results, use less template DNA in the amplification reactions or reduce the number of cycles in the amplification program by 2–4 cycles (i.e., 10/18 or 10/20 cycling).

4. Combine 2.0µl of the prepared loading cocktail and 1.0µl of the PowerPlex® 16 Allelic Ladder Mix. Vortex the allelic ladder mix prior to pipetting.

5. Briefly centrifuge the samples to bring the contents to the bottom of the tubes.

6. Just prior to loading the gel, denature the samples by heating at 95°C for 3 minutes, and immediately chill on crushed ice or in an ice-water bath. Denature the samples just prior to loading the gel.

7. After the 15- to 20-minute pre-run, pause the instrument by selecting "Pause". By pausing the pre-run, the water will continue to circulate, keeping the gel warm during sample loading.

8. Use a 60cc syringe filled with buffer and fitted with a bent 18-gauge needle to flush the urea from the well area.

9. Load 1.5µl of each denatured sample into the respective wells.

   You may need to optimize the volume of sample loaded for individual instruments. We recommend loading volumes of 1.0–2.0µl.

10. Place the lid on the upper buffer chamber, and close the instrument door.

Gel Electrophoresis and Detection

1. After loading, select "Cancel" to stop the pre-run. Make sure that the run time is set at 3 hours, then select "Run" to begin electrophoresis.
2. Monitor the electrophoresis by observing the gel image and status windows.
3. Allow electrophoresis to proceed for 3 hours. The 600-base ILS fragment will have migrated past the laser.
4. Track and extract the gel lanes.

Reuse of Glass Plates

Separate the glass plates, and discard the gel. Clean the glass plates with hot water and a detergent such as 1% Liqui-Nox® detergent. Rinse extremely well with deionized water, and allow the plates to air-dry. Do not scrape the plates with abrasive materials during this process.

Note: Soap and oil may build up on plates, resulting in gel extrusion or hazy background. Soak the plates in 2N HCl for 15 minutes, then rinse thoroughly to remove any buildup.

VI. Data Analysis

VI.A. PowerPlex® Panel and Bin Sets with GeneMapper® *ID*, Version 3.2

To facilitate analysis of the data generated with the PowerPlex® 16 System, we have created panel and bin files to allow automatic assignment of genotypes using GeneMapper® *ID* software, version 3.2. We recommend that users of GeneMapper® *ID* software, version 3.2, complete the *Applied Biosystems GeneMapper®* ID *Software Human Identification Analysis Tutorial* to familiarize themselves with the proper operation of the software. For GeneMapper® *ID* software, version 3.1, users we recommend upgrading to version 3.2.

Getting Started

1. Obtain the proper panel and bin files for use with GeneMapper® *ID* from the Promega web site at: **www.promega.com/geneticidtools/panels_bins/**
2. Enter your contact information, and select "GeneMapper ID version 3.2". Select "Submit".
3. Select the "PowerPlex® Panels & Bin Sets" link, and save the .zip file to your computer.
4. Open the files using the Windows® WinZip program, and save the unzipped files to a known location on your computer.

Importing Panel and Bin Files

These instructions loosely follow the Applied Biosystem GeneMapper® *ID* software tutorial, pages 1–4.

1. Open the GeneMapper® *ID* software, version 3.2.
2. Select "Tools", then "Panel Manager".
3. Highlight the Panel Manager icon in the upper left tile (navigation pane).
4. Select "File", then "Import Panels".
5. Navigate to the saved panel and bin files. Select "Promega_Panels_ID3.2.X.txt", where "X" refers to the most recent version of the panel and bin files. Select "Import".
6. In the navigation pane, highlight the Promega_Panels_ID3.2.X folder that you just imported.
7. Select "File", then "Import Bin Set".
8. Navigate to the saved panel and bin files. Select "Promega_Bins_ID3.2.X.txt", then "Import".
9. At the bottom of the Panel Manager window, select "Apply", then "OK". The panel manager window will close automatically.

VI.B. Creating a Casework Analysis Method with GeneMapper® *ID* Software

These instructions loosely follow the Applied Biosystem GeneMapper® *ID* software tutorial, pages 1–11.

1. Select "Tools", then "GeneMapper Manager".
2. Select the Analysis Methods tab.
3. Select "New", and a new analysis method dialog box will open.
4. Select "HID", and select "OK".

   Note: If you do not see the HID option, you do not have the GeneMapper® *ID* software. Contact Promega Technical Services by e-mail: **genetic@promega.com** for assistance.
5. Enter a descriptive name for the analysis method, such as "PowerPlex16 advanced".
6. Select the Allele tab (Figure 2).

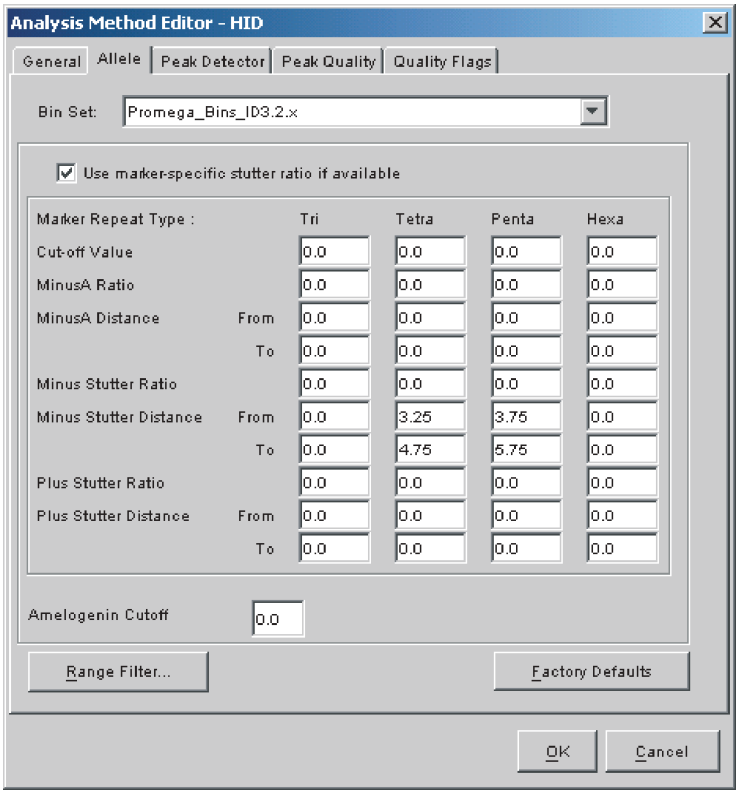

Figure 2. The Allele tab. Select the bin set "Promega_Bins_ID3.2.X.txt", where "X" refers to the most recent version of the bin set.

7. Select the bin set corresponding to the PowerPlex System "Promega_Bins_ID3.2.X", where "X" refers to the most recent version of the bin set.

8. Ensure that the "Use marker-specific stutter ratio if available" box is checked.

9. Enter the values shown in Figure 2 for proper filtering of stutter peaks when using the PowerPlex® 16 System. For an explanation of the proper usage and effects of these settings, refer to the Applied Biosystems user bulletin titled "*Installation Procedures and New Features for GeneMapper* ID *Software 3.2*".

   Note: Some of these settings have been optimized and are different from the recommended settings in the user bulletin.

10. Select the Peak Detector tab. We recommend the settings shown in Figure 3.

    Note: Select full range or partial range for the analysis range. When using a partial range, choose an appropriate analysis range based on the data. Choose a start point after the primer peak and just before the first defined internal lane standard peak to help ensure proper sizing of the internal lane standard.

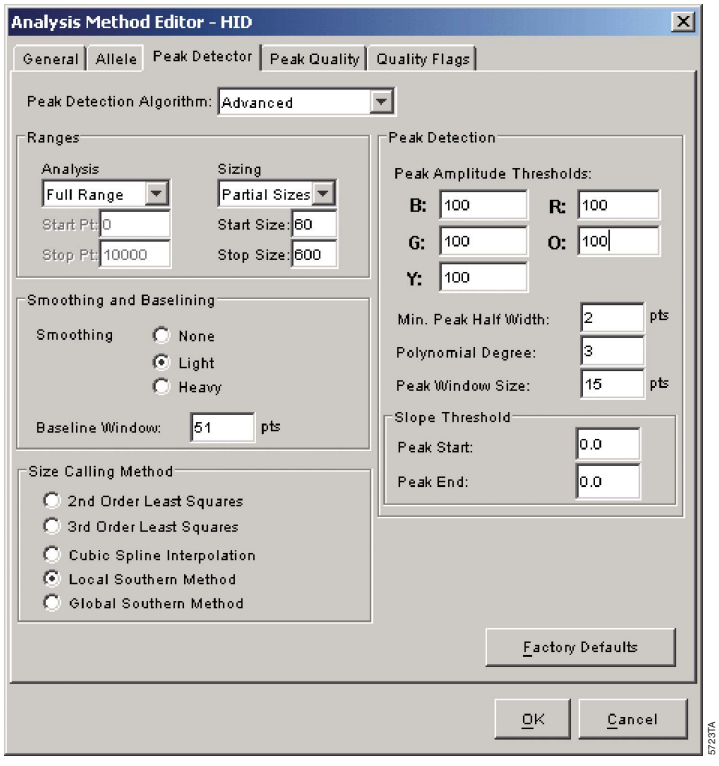

Figure 3. The Peak Detector tab.

11. Select the Peak Quality tab. You may change the settings for peak quality.

    Note: For Steps 11 and 12, see the GeneMapper® *ID* user's manual for more information.

12. Select the Quality Flags tab. You may also change these settings.

13. Select "OK" to save your settings.

Creating a Size Standard

1. Select "Tools", then "GeneMapper Manager".

2. Select the Size Standard tab.

3. Select "New".

4. Select "Basic or Advanced" (Figure 4). The type of analysis method selected must match the type of analysis method created earlier. Select "OK".

5. Enter a detailed name, such as "ILS 600 advanced", in the "Size Standard Editor" (Figure 5).

6. Choose red as the color for the size standard dye.

7. Enter the sizes of the internal lane standard fragments (see Section IX.E, Figure 12).

8. Select "OK".

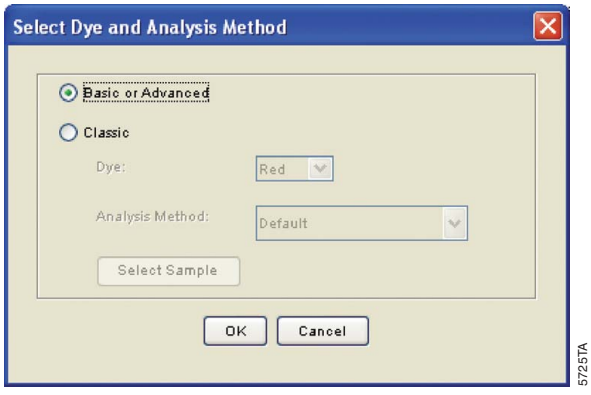

Figure 4. The Select Dye and Analysis Method window.

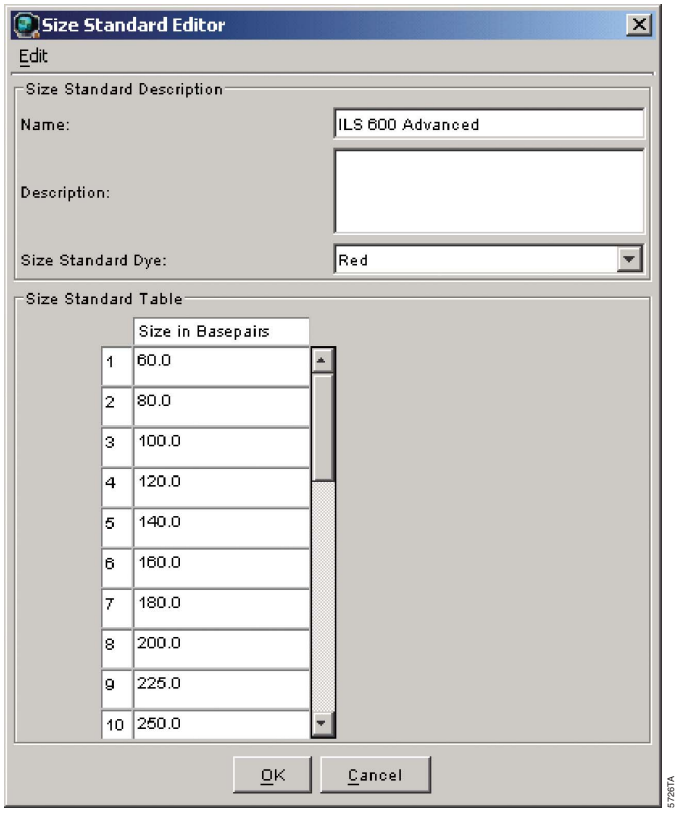

Figure 5. The Size Standard Editor.

Processing Sample Data for Casework

1. Import sample files into a new project as described in the *Applied Biosystems GeneMapper®* ID *Software Human Identification Analysis Tutorial.*
2. In the "Sample Type" column, use the drop-down menu to select "Ladder", "Sample", "Positive Control" or "Negative Control". Every folder in the project must contain at least one ladder that is designated as such for proper genotyping.
3. In the "Analysis Method" column, select the analysis method created previously in the Creating a Casework Analysis Method section.
4. In the "Panel" column, select "PowerPlex_16_ID3.2.X", where "X" refers to the most recent version of the panel files. This is the panel set that was imported in Section VI.A.
5. In the "Size Standard" column, select the size standard that was created in the Creating a Size Standard section.
6. If analyzing data from an ABI PRISM® 310 Genetic Analyzer or an ABI PRISM® 377 DNA Sequencer, ensure that the appropriate matrix file is selected in the "Matrix" column.
7. Select "Analyze" (green arrow button) to start the data analysis.

VI.C. Creating a Databasing or Paternity Analysis Method with GeneMapper® *ID* Software

1. Select "Tools", then "GeneMapper Manager".
2. Select the Analysis Methods tab.
3. Select "New", and a new analysis method dialog box will open.
4. Select "HID", and select "OK".

   Note: If you do not see the HID option, you do not have the GeneMapper® *ID* software. Contact Promega Technical Services by e-mail: **genetic@promega.com** for assistance.
5. Enter a descriptive name for the analysis method, such as "PowerPlex16_20%filter".
6. Select the Allele tab.
7. Select the bin set corresponding to the PowerPlex® System "Promega_Bins_ID3.2.X", where "X" refers to the most recent version of the bin set.
8. Ensure that the "Use marker-specific stutter ratio if available" box is checked.
9. Enter the values shown in Figure 6 for proper filtering of peaks when using the PowerPlex® 16 System. For an explanation of the proper usage and effect of these settings, refer to the Applied Biosystems user bulletin titled "*Installation Procedures and New Features for GeneMapper* ID *Software* 3.2".

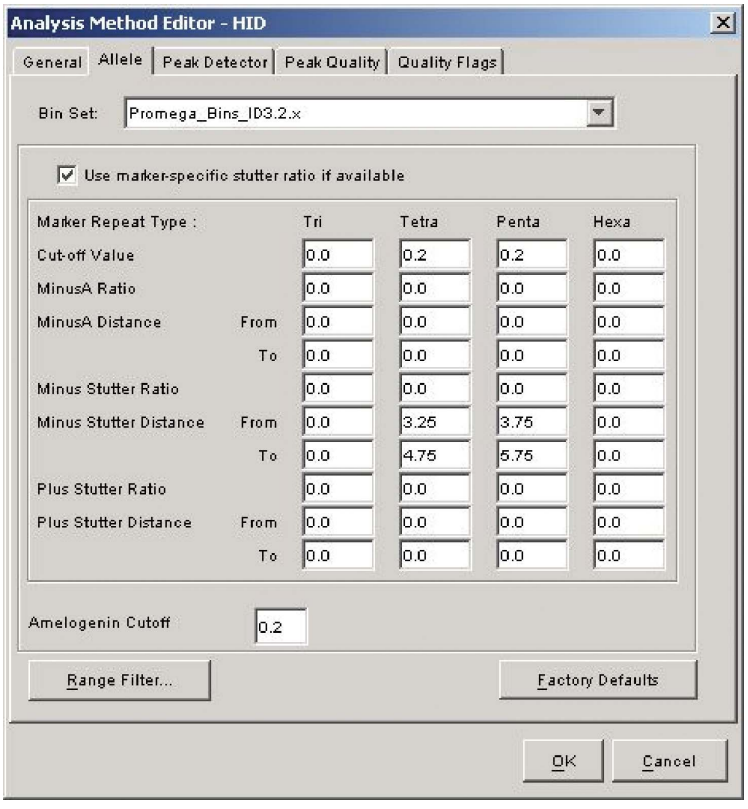

Figure 6. The Allele tab with settings for using a 20% peak filter. Select the bin set "Promega_Bins_ID3.2.X.txt", where "X" refers to the most recent version of the bin set.

Creating a Size Standard

1. Select "Tools", then "GeneMapper Manager".
2. Select the Size Standard tab.
3. Select "New".
4. Select "Basic or Advanced" (Figure 4). The type of analysis method selected must match the type of analysis method created earlier. Select "OK".
5. Enter a detailed name, such as "ILS 600 advanced", in the Size Standard Editor (Figure 5).
6. Choose red as the color for the size standard dye.
7. Enter the sizes of the internal lane standard fragments (see Section IX.E, Figure 12).
8. Select "OK".

Processing Data for Databasing or Paternity Samples

1. Import sample files into a new project as described in the *Applied Biosystems GeneMapper® ID Software Human Identification Analysis Tutorial.*
2. In the "Sample Type" column, use the drop-down menu to select "Ladder", "Sample", "Positive Control" or "Negative Control". Every folder in the project must contain at least one ladder that is designated as such for proper genotyping.
3. In the "Analysis Method" column, select the analysis method created in the Creating a Databasing or Paternity Analysis Method section.
4. In the "Panel" column, select "PowerPlex_16_ID3.2.X", where "X" refers to the most recent version of the panel files. This is the panel set that was imported in Section VI.A.
5. In the "Size Standard" column, select the size standard that was created in the Creating a Size Standard section.
6. If analyzing data from an ABI PRISM® 310 Genetic Analyzer or an ABI PRISM® 377 DNA Sequencer, ensure that the appropriate matrix file is selected in the "Matrix" column.
7. Select "Analyze" (green arrow button) to start the data analysis.

VI.D. Sample Analysis Using the GeneScan® Software and PC Operating Systems

1. Analyze the data using the GeneScan® analysis software.
2. Review the raw data for one or more sample runs. Highlight the sample file name, then under the "Sample" menu, select "raw data". Move the cursor so the crosshair is on the baseline to the right of the large primer peak (before the first internal lane standard peak [red]). Use the X-value number shown at the bottom left of the window for the start position in the analysis parameters.
3. The recommended analysis parameters are shown in Figure 7.
4. The analysis parameters can be saved in the Params folder; in most installations this is located at:
   C:\AppliedBio\Shared\Analysis\Sizecaller\Params
5. Apply the stored analysis parameters file to the samples.
6. Assign a new size standard. Select a sample file, and highlight the arrow next to size standard. Select "define new". Assign the size standard peaks as shown in Figure 12 in Section IX.E. Store the size standard in the Size Standards folder at:
   C:\AppliedBio\Shared\Analysis\Sizecaller\SizeStandards
7. Apply the size standard file to the samples, then analyze the sample files. See Section VI.F for additional information on the use of the PowerTyper™ 16 Macro (Release 2.0) and Genotyper® software.

Notes:

1. Peak heights outside the linear range of the instrument may generate artifact peaks due to instrument saturation (i.e., overloading the sample). Bleedthrough (pull-ups) from one color to another may be observed. Saturated signal may also appear as two peaks (split peak).
2. If the peak heights are not within the linear range of detection of the instrument, the ratio of stutter peaks to real allele peaks increases, and allele designations become difficult to interpret. The balance of the peak heights may also appear less uniform.
3. There may be variation between instruments regarding the relative fluorescence levels detected using the same sample. Furthermore, different instruments vary in the relative efficiency of color detection, affecting the dye color-to-dye color balance.

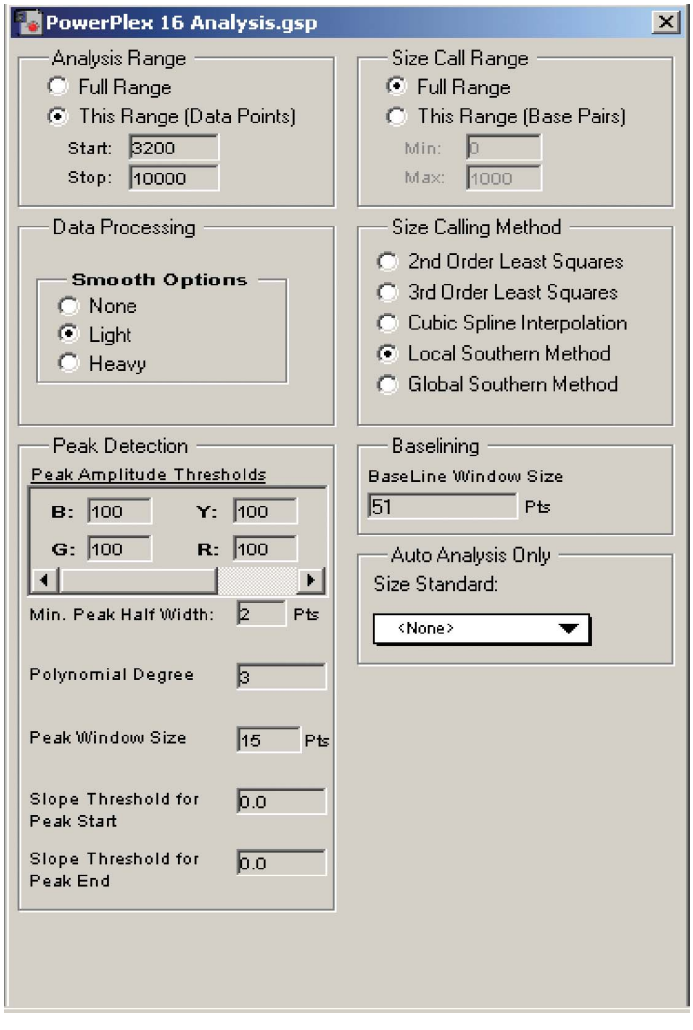

Figure 7. The analysis parameters window. The start point of the analysis range, which will vary, is defined in Section VI.D, Step 2.

VI.E. Sample Analysis Using the GeneScan® Software and Macintosh® Operating Systems

1. Analyze the data using the GeneScan® analysis software.
2. Review the raw data for one or more sample runs. Highlight the sample file name, then under the "Sample" menu, select "raw data". Move the cursor so the crosshair is on the baseline to the right of the large primer peak (before the first internal lane standard peak [red]). Use the X-value number shown at the bottom left of the window for the start position in the analysis parameters.
3. The recommended analysis parameters are:

Analysis Range	Start: Defined in Step 2 Stop: 10,000
Data Processing	Baseline: Checked Multicomponent: Checked Smooth Options: Light[1]
Peak Detection	Peak Amplitude Thresholds[2]: B: Y: G: R: Min. Peak Half Width: 2pts
Size Call Range	Min: 60 Max: 600
Size Calling Method	Local Southern Method
Split Peak Correction	None

[1]Smooth options should be determined by individual laboratories. Occasionally the TH01 alleles 9.3 and 10 will not be distinguished using heavy smoothing.

[2]The peak amplitude thresholds are the minimum peak heights that the software will call as a peak. Values for the peak amplitude thresholds are usually 50–200RFU and should be determined by individual laboratories.

4. The analysis parameters can be saved in the Params folder.
5. Apply the stored analysis parameters file to the samples.
6. Assign a new size standard. Select a sample file, highlight the arrow next to size standard, then select "define new". Assign the size standard peaks as shown in Figure 12 in Section IX.E. Store the size standard in the Size Standards folder.
7. Apply the size standard file to the samples, then analyze the sample files. See Section VI.F for additional information on the use of the PowerTyper™ 16 Macro (Release 2.0) and Genotyper® software.

   For additional information regarding the GeneScan® analysis software, refer to the *GeneScan® Analysis Software User's Manual*.

Notes:

1. Peak heights outside the linear range of the instrument may generate artifact peaks due to instrument saturation (i.e., overloading the sample). Bleedthrough (pull-ups) from one color to another may be observed. Saturated signal may also appear as two peaks (split peak).
2. If the peak heights are not within the linear range of detection of the instrument, the ratio of stutter peaks to real allele peaks increases, and allele designations become difficult to interpret. The balance of the peak heights may also appear less uniform.
3. There may be variation between instruments regarding the relative fluorescence levels detected using the same sample. Furthermore, different instruments vary in the relative efficiency of color detection, affecting the dye color-to-dye color balance.

VI.F. Sample Analysis Using the Genotyper® Software and PowerTyper™ 16 Macro

To facilitate analysis of the data generated with the PowerPlex® 16 System, we have created a file to allow automatic assignment of genotypes using the Genotyper® software. After samples are amplified, detected using the ABI PRISM® 310 or 3100 Genetic Analyzer (using data collection software, version 1.0.1 or 1.1) or ABI PRISM® 377 DNA Sequencer, and analyzed using the GeneScan® analysis software, the sample files can be imported into the Genotyper® program and analyzed using the PowerTyper™ 16 Macro (Release 2.0).

The PowerTyper™ 16 Macro (Release 2.0) is available upon request from Promega. The PowerTyper™ 16 Macro (Release 2.0) is provided on the PowerTyper™ Macros CD-ROM (Cat.# DG3470). The PowerTyper™ Macros can also be downloaded from the Promega web site at: **www.promega.com/geneticidtools/**

The PowerTyper™ 16 Macro (Release 2.0) is used in conjunction with Macintosh® Genotyper® software, version 2.5, and Windows NT® Genotyper® software, version 3.6, or later. The Genotyper® software must be installed on your computer before the PowerTyper™ 16 Macro (Release 2.0) can be used.

Be certain the "sample info" (Macintosh® computers) or "color info" (Windows NT® operating systems) column for each lane containing the allelic ladder mix contains the word "ladder". The macro uses the word "ladder" to identify the sample file(s) containing allelic ladder. Sample info can be added or modified after importing into the PowerTyper™ Macro. Highlight the sample, then select "show dye/lanes window" under "Views".

1. Transfer the PowerTyper™ 16 Macro (Release 2.0) from the PowerTyper™ Macros CD-ROM (Cat.# DG3470) to a designated location on your computer hard drive. Alternatively, download the PowerTyper™ 16 Macro (Release 2.0) from the Promega web site.
2. Open the Genotyper® software, then the PowerTyper™ 16 Macro (Release 2.0). For questions about the Genotyper® software, refer to the *Genotyper® Analysis Software User's Manual*.

3. Under "File", select "Import", and import the GeneScan® project or sample files to be analyzed. Import the blue, yellow, green and red dye colors.

   Note: To select the dye colors to be imported, select "Set Preferences" under the "Edit" menu.

4. Double-click on the Check ILS macro. The macros are listed at the bottom left corner of the active window. A plots window will be displayed to show the internal lane standard (i.e., ILS 600) in the red dye color. Scroll down to view, and confirm that the internal lane standard fragment sizes are correct. If necessary, re-analyze samples using the GeneScan® software and redefine the internal lane standard fragments.

   Note: The software uses one ladder sample for determining allele sizes. The macro uses the first ladder sample imported for allele designations.

5. For casework, double-click on the POWER macro. The POWER macro identifies the alleles in the ladder sample and calculates offsets for all of the loci. This process may take several minutes. When completed, a plots window will open to display the allelic ladders (i.e., Penta E, D18S51, D21S11, TH01 and D3S1358).

   Alternatively, for databasing or paternity, double-click on the POWER 20% Filter macro. This macro has a higher level of filtering than the standard POWER macro to reduce the need for manual editing of peak labels. The POWER 20% Filter should not be used if mixtures may exist.

   In general, the allelic ladders contain fragments of the same lengths as many known alleles for the locus. The allelic ladder sizes and repeat units are listed in Table 5 (Section IX.B). Analysis using GeneScan® analysis software and Genotyper® software allows allele determination by comparing amplified sample fragments with allelic ladders and internal lane standards. When using an internal lane standard, the calculated lengths of the allelic ladder components may differ from those listed in the table. This is due to differences in migration resulting from sequence differences between the allelic ladder fragments and the internal size standard and is not a matter of concern.

6. Double-click on the Allelic Ladders macro. A plots window will open to display the blue (fluorescein) dye allelic ladders (i.e., Penta E, D18S51, D21S11, TH01 and D3S1358), the green (JOE) dye allelic ladders (i.e., Penta E, CSF1PO, D16S539, D7S820, D13S317 and D5S818) and the yellow (TMR) dye allelic ladders (i.e., FGA, TPOX, D8S1179, vWA and Amelogenin). Confirm that the correct allele designations were assigned to the allelic ladders (Figure 10 in Section VI.H).

   Note: The software uses one ladder sample for determining the allele sizes. The macro uses the first ladder sample imported for allele designations. If the POWER macro is run a second time, the software will use the second ladder; if the POWER macro is run a third time, the software will use the third ladder, etc., until all ladders in the project are used. If an allelic ladder fails to be analyzed or if many off-ladder alleles are found in the samples, the samples should be re-analyzed using another ladder from the project.

7. Double-click on the Display Fluorescein Data macro to display the blue dye for all the sample injections/lanes. Scroll down to observe and edit as needed.
8. Double-click on the Display TMR Data macro to display the yellow dye for all sample injections/lanes. Scroll down to observe and edit as needed.
9. Double-click on the Display JOE Data macro to display the green dye for all sample injections/lanes. Scroll down to observe and edit as needed.
10. Create the appropriate table by selecting the "PowerTable", "Make Allele Table" or "Make CODIS Table" macro. The three available table formats are shown below. The PowerTable option allows up to four alleles per sample file. Additional information such as low peak signal or high peak signal is also included. The Allele Table and CODIS Table options include only two alleles per locus. If more than two alleles are present at a locus, the smallest alleles identified are included. The Allele Table format displays the categories (loci) in columns, while the CODIS table format displays the categories in rows. These tables can be customized to fit needs. To save data in tables, go to the Table drop-down menu, highlight "Export to File..." and save the file with the desired name and location. The saved file can be viewed and analyzed using Microsoft® Excel.

PowerTable Format

Sample Info	Sample Comment	Category	Peak 1	Peak 2	Peak 3	Peak 4	Over-flow	Low Signal	Satura-tion	Edited Label	Edited Row

Allele Table Format

Sample Info	Category Allele 1	Category Allele 2	Category Allele 1	Category Allele 2	Category Allele 1	Category Allele 2	Category Allele 1	Category Allele 2

CODIS Table Format

Sample Info	Category	Peak 1	Peak 2

11. Save the analyzed data, go to the "File" menu and select "Save as".

The PowerTyper™ Macro is a Genotyper® file and can be overwritten if "Save" is used instead of "Save as".

VI.G. Controls

1. Observe the results for the negative control. The negative control should be devoid of amplification products.
2. Observe the results for the 9947A positive control DNA. Compare the control DNA allelic repeat sizes with the locus-specific allelic ladder. The expected 9947A DNA allele designations for each locus are listed in Table 6 (Section IX.B).

VI.H. Results

Representative results of the PowerPlex® 16 System are shown in Figure 8. The PowerPlex® 16 Allelic Ladder Mix is shown in Figure 9.

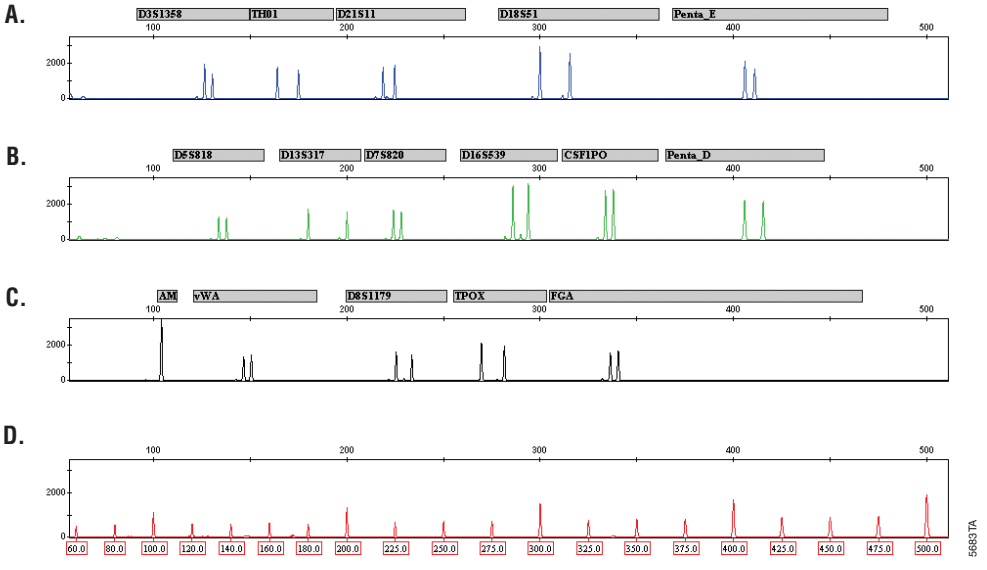

Figure 8. The PowerPlex® 16 System. A single template DNA (1.0ng) was amplified using the PowerPlex® 16 10X Primer Pair Mix. The amplification products were mixed with the Internal Lane Standard 600 and analyzed with an Applied Biosystems 3130 Genetic Analyzer using a 3kV, 5-second injection. The results were analyzed using GeneMapper® *ID* software, version 3.2. **Panel A.** An electropherogram showing the peaks of the fluorescein-labeled loci: D3S1358, TH01, D21S11, D18S51 and Penta E. **Panel B.** An electropherogram showing the peaks of the JOE-labeled loci: D5S818, D13S317, D7S820, D16S539, CSF1PO and Penta D. **Panel C.** An electropherogram showing the peaks of the TMR-labeled loci: Amelogenin, vWA, D8S1179, TPOX and FGA. **Panel D.** An electropherogram showing the 60bp to 500bp fragments of the Internal Lane Standard 600.

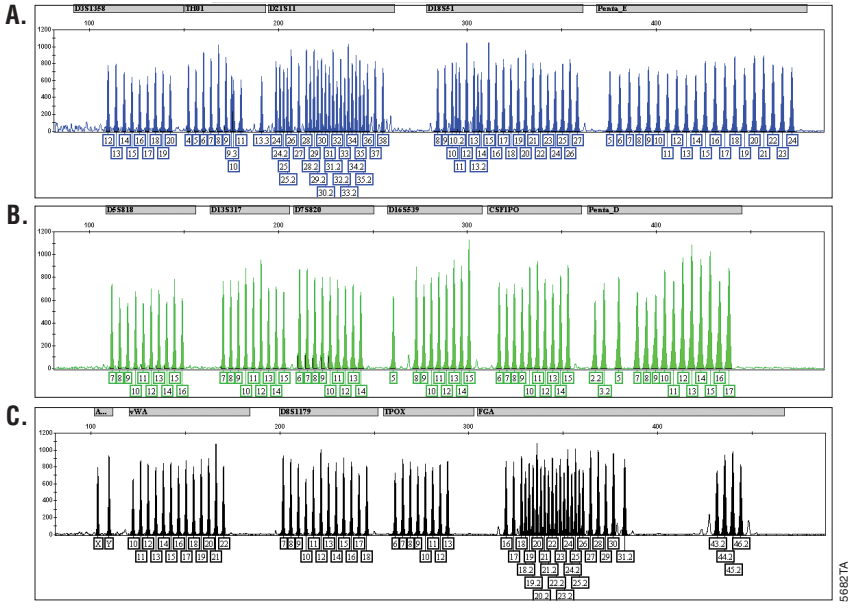

Figure 9. The PowerPlex® 16 Allelic Ladder Mix. The PowerPlex® 16 Allelic Ladder Mix was analyzed with an Applied Biosystems 3130 Genetic Analyzer using a 3kV, 5-second injection. The sample file was analyzed with the GeneMapper® *ID* software, version 3.2, and the PowerPlex® 16 panel and bin files. **Panel A.** The fluorescein-labeled allelic ladder components and their allele designations. **Panel B.** The JOE-labeled allelic ladder components and their allele designations. **Panel C.** The TMR-labeled allelic ladder components and their allele designations.

Artifacts and Stutter

Stutter bands are a common amplification artifact associated with STR analysis. Stutter products are often observed one repeat unit below the true allele peak and, occasionally, two repeat units smaller or one repeat unit larger than the true allele peak. Frequently, alleles with a greater number of repeat units will exhibit a higher percent stutter. The pattern and intensity of stutter may differ slightly between primer sets for the same loci. Stutter was determined and published as part of the validation of PowerPlex® 16 System (9).

In addition to stutter peaks, other artifact peaks can be observed at some of the PowerPlex® 16 System loci. Low-level products can be seen in the n–2 and n+2 positions (two bases below and above the true allele peak, respectively) with some loci such as D21S11. Samples may show low-level artifacts in the noncalling regions between the D7S820 and D13S317 allele ranges and between the D3S1358 and TH01 allele ranges. Occasionally an off-ladder artifact can be seen in the 270–271bp position in the JOE dye channel. One or more extra peaks that are not directly related to amplification may be observed at positions 8–26 bases smaller than TPOX alleles and 6–21 bases smaller than vWA alleles. These extra peaks occur when the amplified peaks are particularly intense (high signal level or template amount), formamide, polymer or capillary was of poor quality or denaturation was ineffective. Please see Section VII for more information about how to minimize these artifacts.

VII. Troubleshooting

For questions not addressed here, please contact your local Promega Branch Office or Distributor. Contact information available at: **www.promega.com**. E-mail: **genetic@promega.com**

VII.A. Amplification and Fragment Detection

Symptoms	Causes and Comments
Faint or absent allele peaks	Impure template DNA. Because of the small amount of template used, this is rarely a problem. Depending on the DNA extraction procedure used and the sample source, inhibitors may be present in the DNA sample.
	Insufficient template. Use the recommended amount of template DNA.
	Insufficient template. Low-copy-number (LCN) analysis using capillary electrophoresis may benefit from reducing competing charged particles during injection. This can be accomplished with post-PCR cleanup or desalting, lower-conductivity formamide or reduced amounts of ILS 600. In-house validation should be performed for any of these methods.
	Insufficient enzyme activity. Use the recommended amount of AmpliTaq Gold® DNA polymerase. Check the expiration date on the tube label.
	Incorrect amplification program. Confirm the amplification program.
	High salt concentration or altered pH. If the DNA template is stored in TE buffer that is not pH 8.0 or contains a higher EDTA concentration, the DNA volume should not exceed 20% of the total reaction volume. Carryover of K^+, Na^+, Mg^{2+} or EDTA from the DNA sample can negatively affect PCR. A change in pH may also affect PCR. Store DNA in TE^{-4} buffer (10mM Tris HCl [pH 8.0], 0.1mM EDTA) or nuclease-free water.
	Thermal cycler, plate or tube problems. Review the thermal cycling protocols in Section IV.B. We have not tested other reaction tubes, plates or thermal cyclers. Calibrate the thermal cycler heating block, if necessary.
	Primer concentration was too low. Use the recommended primer concentration. Mix the 10X PowerPlex® 16 Primer Pair for 15 seconds using a vortex mixer before use.
	Poor capillary electrophoresis injection (ILS 600 peaks also affected). Re-inject the sample. Check the syringe for leakage. Check the laser power.
	Poor-quality formamide was used. Use only Hi-Di™ formamide when running samples.

Symptoms	Causes and Comments
Extra peaks visible in one or all of the color channels	Contamination with another template DNA or previously amplified DNA. Cross-contamination can be a problem. Use aerosol-resistant pipette tips, and change gloves regularly.
	Samples were not completely denatured. Heat denature the samples for the recommended time, and cool on crushed ice or in an ice-water bath immediately prior to loading the gel or capillary.
	Artifacts of STR amplification. PCR amplification of STR systems sometimes generates artifacts that appear as faint peaks one repeat unit smaller than the allele. Stutter band peak heights can be high if the samples are overloaded.
	Artifacts of STR amplification. PCR amplification of STR systems can result in artifacts that appear as peaks one base smaller than the allele due to incomplete addition of the 3´ A residue. Be sure to perform the 30-minute extension step at 60°C after thermal cycling (Section IV.B).
	High background. Load less amplification product or decrease the injection time. See Section V.
	Capillary electrophoresis (CE)-related artifacts ("spikes"). Minor voltage changes or urea crystals passing by the laser can cause "spikes" or unexpected peaks. Spikes sometimes appear in one color but often are easily identified by their presence in more than one color. Re-inject the samples to confirm.
	CE-related artifacts (contaminants). Contaminants in the water used with the instrument or to dilute the 10X genetic analyzer buffer may generate peaks in the blue and green dye colors. Use autoclaved water, change vials and wash buffer reservoir.
	Excessive amount of DNA. Amplification of >2ng template can result in a higher number of stutter bands. Use less template DNA, or reduce the number of cycles in the amplification program by 2–4 cycles (10/20 or 10/18 cycling).
	Pull-up or bleedthrough. Pull-up can occur when peak heights are too high or if a poor or incorrect matrix has been applied to the samples. • For the ABI PRISM® 310 Genetic Analyzer and 377 DNA Sequencer, generate a new matrix, and apply it to the samples. For the ABI PRISM® 3100 and 3100-*Avant* Genetic Analyzers and Applied Biosystems 3130 and 3130*xl* Genetic Analyzers, perform a new spectral calibration and re-run the samples. • Sensitivities of instruments may vary. Optimize the injection or gel loading conditions. See Section V.
	Long-term storage of amplified sample in formamide can result in degradation. Repeat preparation of samples using fresh formamide.
	The CE polymer was beyond its expiration date, or polymer was stored at room temperature for more than one week.
	Maintain instrumentation on a daily or weekly basis, as recommended by the manufacturer.

Symptoms	Causes and Comments
Allelic ladder not running the the same as the sample	Allelic ladder and primer pair mix were not compatible. Ensure that the allelic ladder is from the same kit as the primer pair mix.
	Buffer incompatibility. Samples were diluted in the wrong buffer. Use Gold ST★R 1X Buffer to dilute samples.
	Poor-quality formamide. Use only Hi-Di™ formamide when analyzing samples.
	Be sure the allelic ladder and samples are from the same instrument run.
	Migration of samples changed slightly over the course of a CE run with many samples. This may be due to changes in temperature or the CE column over time. Use a different injection of the allelic ladder to determine sizes.
	Poor injection of allelic ladder. Include more than one ladder per instrument run.
Peak height imbalance	Excessive amount of DNA. Amplification of >1ng of template can result in an imbalance with yields with smaller loci showing more product than the larger loci. Use less template, or reduce the number of cycles in the amplification program by 2–4 cycles (10/20 or 10/18 cycling) to improve locus-to-locus balance. **Note:** Dilution of overamplified samples can result in dropout of larger loci.
	Use of FTA® paper. Results may be similar to those obtained with excess amounts of DNA template. Reduce the number of cycles in the amplification program by 2–4 cycles (10/20 or 10/18 cycling) to improve locus-to-locus balance.
	Degraded DNA sample. DNA template is degraded, and the larger loci show diminished yield. Repurify the template DNA.
	Insufficient template DNA. Use the recommended amount of template DNA. Stochastic effects can occur when amplifying low amounts of template.
	Miscellaneous balance problems. Thaw the 10X Primer Pair Mix and Gold ST★R 10X Buffer completely, and vortex for 15 seconds before using. Do not centrifuge the 10X Primer Pair Mix after mixing. Calibrate thermal cyclers and pipettes routinely. Using a 59°C annealing temperature instead of 60°C has been shown to improve balance in some instances.
	Impure template DNA. Inhibitors that may be present in forensic samples can lead to allele dropout or imbalance.

VII.B. GeneMapper® *ID* Analysis Software

Symptoms	Causes and Comments
Alleles not called	To analyze samples with GeneMapper® *ID* software, the analysis parameters and size standard must both have "Basic or Advanced" as the analysis type. If they are different, an error is obtained (Figure 10).
	An insufficient number of ILS 600 fragments was defined. Be sure to define at least one ILS 600 fragment smaller than the smallest sample peak and at least one ILS 600 fragment larger than the largest sample peak.
	Run was too short, and larger peaks in ILS were not captured. Not all of the ILS 600 peaks defined in the size standard were detected during the run. • Create a new size standard using the internal lane standard fragments present in the sample. • Re-run the samples using a longer run time.
Off-ladder alleles	An allelic ladder from a different run than the samples was used. Re-analyze the samples with an allelic ladder from the same run.
	The GeneMapper® *ID* software requires that the allelic ladder be imported from the same folder as the sample. Be sure that the allelic ladder is in the same folder as the sample. Create a new project and re-analyze, as described in Section VI.B or VI.C.
	Panel file selected for analysis was incorrect for the STR system used. Assign correct panel file that corresponds to the STR system used for amplification.
	The allelic ladder was not identified as an allelic ladder in the sample type column.
	The wrong analysis type was chosen for the analysis method. Be sure to use the HID analysis type.
	The internal lane standard was not properly identified in the sample. Manually redefine the sizes of the size standard fragments in the sample.
Size standard not called correctly (Figure 11)	Starting data point was incorrect for the partial range chosen in Section VI.B. Adjust the starting data point in the analysis method. Alternatively, use a full range for the analysis.
	Extra peaks in advanced mode size standard. Open the size match editor. Highlight the extra peak, select "Edit" and select "delete size label". Select "auto adjust sizes".
	Run was too short, and larger peaks in ILS were not captured. Not all of the ILS 600 peaks defined in the size standard were detected during the run. • Create a new size standard using the internal lane standard fragments present in the sample. • Re-run the samples using a longer run time.
Peaks in size standard missing	If peaks are below threshold, decrease the peak threshold in the analysis method for the red channel to include peaks.
	If peaks are low-quality, redefine the size standard for the sample to skip these peaks.

Symptoms	Causes and Comments
Error message: "Either panel, size standard, or analysis method is invalid"	The size standard and analysis method were not in the same mode ("Classic" vs. "Basic or Advanced"). Be sure both files are set to the same mode, either Classic or Basic or Advanced mode.
No alleles called, but no error message appears	Panel was not selected for sample. In "Panel" column, select the appropriate panel set for the STR system that was used.
	No size standard was selected. In the size standards column, be sure to select the appropriate size standard.
	Size standard was not correctly defined or size peaks were missing. Redefine size standard to include only peaks present in your sample. Terminating analysis early or using short run times will cause larger ladder peaks to be missing. This will cause your sizing quality to be flagged as "red", and no allele sizes will be called.
Error message: "Both the Bin Set used in the Analysis Method and the Panel must belong to the same Chemistry Kit".	The bin set assigned to the analysis method may have been deleted. In the GeneMapper® Manager, select the Analysis Methods tab, and open the analysis method of interest. Select the Alleles tab, and select an appropriate bin set.
	The wrong bin set was chosen in the analysis method Allele tab. Be sure to choose the appropriate bin set, as shown in Figure 2.
Significantly raised baseline	• Poor spectral calibration for the ABI PRISM® 3100 and 3100-*Avant* Genetic Analyzers and Applied Biosystems 3130 and 3130*xl* Genetic Analyzers. Perform a new spectral calibration and re-run the samples. • Poor matrix for the ABI PRISM® 310 Genetic Analyzer. Re-run and optimize the matrix.
	Use of Classic mode analysis method. Use of Classic mode analysis on samples can result in baselines with more noise than those analyzed using the Basic or Advanced mode analysis method. Advanced mode analysis methods and size standards are recommended.
Red bar appears during analysis of samples, and the following error message appears when data are displayed: "Some selected sample(s) do not contain analysis data. Those sample(s) will not be shown".	If none of the samples had matrices applied when run on the ABI PRISM® 310 Genetic Analyzer, no data will be displayed. Apply a matrix file during analysis in the GeneMapper® *ID* software and re-analyze.
Error message after attempting to import panel and bin files: "Unable to save panel data: java.SQLEException: ORA-00001: unique constraint (IFA.CKP_NNN) violated".	There was a conflict between different sets of panel and bin files. Delete all panel and bin sets, and re-import files in a different order.

VII.B. GeneMapper® *ID* Analysis Software (continued)

Symptoms	Causes and Comments
Peaks in the allelic ladder are labeled off-ladder	GeneMapper® *ID* software was not used or microsatellite analysis settings were used instead of HID analysis settings. GeneMapper® software does not use the same algorithms as GeneMapper® *ID* software and cannot correct for sizing differences using the allelic ladder. Promega recommends using GeneMapper® *ID* software to analyze PowerPlex® reactions. If using GeneMapper® *ID* software, version 3.1 or 3.2, be sure that the analysis method selected is an HID method. This can be verified by opening the analysis method using the GeneMapper® Manager, then selecting the General tab. The analysis type cannot be changed. If the method is not HID, it should be deleted and a new analysis method created.

VII.C. PowerTyper™ 16 Macro

Symptoms	Causes and Comments
File does not open on your computer	Genotyper® software was not installed. Be certain that the Genotyper® software, version 2.5 (Macintosh®) or version 3.6 or higher (Windows NT®), is installed.
	Incorrect version of Genotyper® software. The PowerTyper™ 16 Macro will not work with Genotyper® software versions prior to version 2.5.
	The CD-ROM may have been damaged during shipment. Contact Technical Services by e-mail: **genetic@promega.com**
	The file was corrupted during download or transfer. Download the file again, or obtain the file on CD-ROM.
Error message: "Could not complete the "Run Macro" command because no dye/lanes are selected"	Allelic ladder sample files were not identified. Be certain the "sample info" or "color info" column for each lane containing the PowerPlex® 16 Allelic Ladder Mix contains the word "ladder". The macro uses the word "ladder" to identify the sample files containing the allelic ladder.
	All four dye colors were not imported. For Genotyper® software, versions 2.5 and 3.5 or higher, set the preferences (under "Edit") to import the blue, green, yellow and red colors.
Error message: "Could not complete the "Run Macro" command because the labeled peak could not be found"	The peak heights for one or more of the alleles in the allelic ladder sample file were below 150RFU. The allelic ladder categories are defined as having a minimum peak height of 150RFU. If the peak heights of the ladder alleles are below 150RFU, the software will not be able to locate the allele peak. Re-run the allelic ladder using more sample or longer injection time to assure peak heights above 150RFU.
	CE spikes in the allelic ladder sample were identified as alleles by the macro. Use a different injection of the allelic ladder.
	TH01 9.3 and 10 alleles were not separated when using heavy smoothing in the GeneScan® analysis parameters. Use light smoothing in the GeneScan® analysis parameters.
	Allelic ladder data was not compatible with the PowerTyper™ file used. Confirm that the PowerTyper™ Macro file matches the allelic ladder being used.

Symptoms	Causes and Comments
Error message: "Could not complete the "Run Macro" command because the labeled peak could not be found" (continued)	The base-pair size of the alleles in the allelic ladder are outside of the defined category range. Be sure the internal lane standard fragments are correctly sized. Redefine the internal lane standard fragments, and re-analyze the sample using GeneScan® software. Compare the size of the smallest allele in the allelic ladder with the base-pair size and range listed in the categories for the same alleles. If necessary, increase the category start range (in the category window) to greater than ±6bp, and save the macro under a new name.
	Allelic ladder peaks were too high, causing stutter peaks to be called as allele peaks. Use a shorter injection time, decrease the amount of allelic ladder used or re-analyze the allelic ladder sample using increased peak amplitude thresholds in the GeneScan® analysis parameters.
The plots window or allele table does not display all data	The macros were not run in the proper order. Use the POWER or POWER 20% Filter macro option.
	All four dye colors were not imported. For Genotyper® software, versions 2.5 and 3.5 or higher, set the preferences (under "Edit") to import the blue, green, yellow and red colors.
The Check ILS macro displays an empty plot window	All four dye colors were not imported. For Genotyper® software, versions 2.5 and 3.5 or higher, set the preferences (under "Edit") to import the blue, green, yellow and red colors.
Off-ladder peaks	Migration of samples changed slightly over the course of a CE run with many samples. This may be due to changes in temperature or the CE column over time. Use a different injection of the allelic ladder to determine sizes in the PowerTyper™ 16 Macro (Release 2.0). Do not use the first injection on a new column for the ladder sample.
	The base-pair size of the alleles was incorrect because incorrect fragment sizes were assigned to the internal lane standard. Confirm that the internal lane standard fragment sizes are assigned correctly. Re-analyze the sample using GeneScan® software, and redefine the internal lane standard fragments.

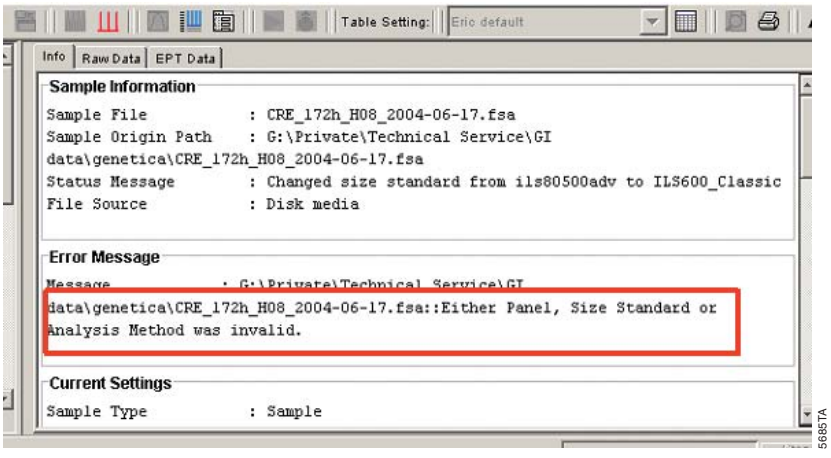

Figure 10. The error message that appears in the GeneMapper® *ID* software when the analysis parameters and the size standard have different analysis types.

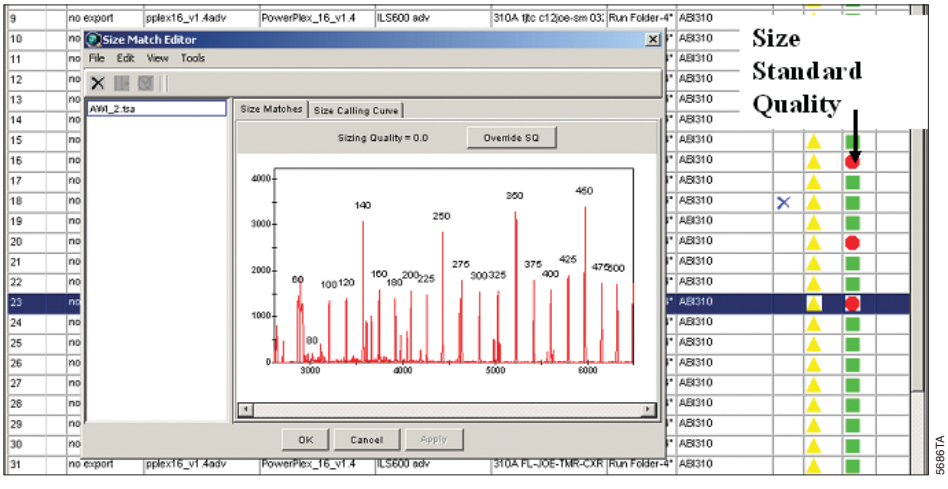

Figure 11. An example showing improper assignment of size standard fragments in the GeneMapper® *ID* software.

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Additional STR references can be found at: **www.promega.com/geneticidentity/**

IX. Appendix

IX.A. Advantages of STR Typing

STR typing is more tolerant of degraded DNA templates than other typing methods because the amplification products are less than 500bp long, much smaller than the material detected using AMP-FLP (13) or VNTR (14) analysis. STR typing is also amenable to a variety of rapid DNA purification techniques, which are compatible with PCR but do not provide enough DNA of appropriate quality for Southern blot-based analyses.

Amplification products generated with Promega STR products are generally of discrete and separable lengths. This allows the construction of allelic ladders containing fragments of the same lengths as several or all known alleles for each locus. Visual or software-based comparison between the allelic ladder and amplified samples of the same locus allows rapid and precise assignment of alleles. Results obtained using the PowerPlex® 16 System can be recorded in a digitized format, allowing direct comparison with stored databases. Population analyses do not require the use of arbitrarily defined fixed bins for population data (15).

IX.B. Advantages of Using the Loci in the PowerPlex® 16 System

The loci included in the PowerPlex® 16 System (Tables 4 and 5) have been selected because they satisfy the needs of several major standardization bodies throughout the world. For example, the United States Federal Bureau of Investigation (FBI) has selected 13 STR core loci for typing prior to searching or including (submitting) samples in CODIS (Combined DNA Index System), the U.S. national database of convicted offender profiles. The PowerPlex® 16 System amplifies all CODIS core loci in a single reaction.

The PowerPlex® 16 System also contains two low-stutter, highly polymorphic pentanucleotide repeat loci, Penta E and Penta D. These additional loci add significantly to the discrimination power of the system, making the PowerPlex® 16 System a single-amplification system with a power of exclusion sufficient to resolve paternity disputes definitively. In addition, the extremely low level of stutter seen with Penta E and Penta D makes them ideal loci for evaluation of DNA mixtures often encountered in forensic casework. Finally, the Amelogenin locus is included in the PowerPlex® 16 System to allow gender identification of each sample. Table 6 lists the PowerPlex® 16 System alleles revealed in commonly available standard DNA templates.

We have carefully selected STR loci and primers to avoid or minimize artifacts, including those associated with *Taq* DNA polymerase, such as repeat slippage and terminal nucleotide addition. Repeat slippage (16,17), sometimes called "n–4 bands", "stutter" or "shadow bands", is due to the loss of a repeat unit during DNA amplification, somatic variation within the DNA in sample material, or both. The amount of this artifact observed depends primarily on the locus and the DNA sequence being amplified.

Terminal nucleotide addition (18,19) occurs when *Taq* DNA polymerase adds a nucleotide, generally adenine, to the 3´ ends of amplified DNA fragments in a template-independent manner. The efficiency with which this occurs varies with different primer sequences. Thus, an artifact band one base shorter than expected (i.e., missing the terminal addition) is sometimes seen. We have modified primer sequences and added a final extension step of 60°C for 30 minutes (20) to the amplification protocols to provide conditions for essentially complete terminal nucleotide addition when recommended amounts of template DNA are used.

The presence of microvariant alleles (alleles differing from one another by lengths other than the repeat length) complicates interpretation and assignment of alleles. There appears to be a correlation between a high degree of polymorphism, a tendency for microvariants and increased mutation rate (21,22). Thus, FGA and D21S11 display numerous, relatively common microvariants. For reasons yet unknown, the highly polymorphic Penta E locus does not display frequent microvariants (Table 5).

Table 4. The PowerPlex® 16 System Locus-Specific Information.

STR Locus	Label	Chromosomal Location	GenBank® Locus and Locus Definition	Repeat Sequence[1] 5´→ 3´
Penta E	FL	15q	NA	AAAGA
D18S51	FL	18q21.3	HUMUT574	AGAA (23)
D21S11	FL	21q11–21q21	HUMD21LOC	TCTA Complex (23)
TH01	FL	11p15.5	HUMTH01, human tyrosine hydroxylase gene	AATG (23)
D3S1358	FL	3p	NA	TCTA Complex
FGA	TMR	4q28	HUMFIBRA, human fibrinogen alpha chain gene	TTTC Complex (23)
TPOX	TMR	2p23–2pter	HUMTPOX, human thyroid peroxidase gene	AATG
D8S1179	TMR	8q	NA	TCTA Complex (23)
vWA	TMR	12p12–pter	HUMVWFA31, human von Willebrand factor gene	TCTA Complex (23)
Amelogenin[2]	TMR	Xp22.1–22.3 and Y	HUMAMEL, human Y chromosomal gene for Amelogenin-like protein	NA
Penta D	JOE	21q	NA	AAAGA
CSF1PO	JOE	5q33.3–34	HUMCSF1PO, human c-fms proto-oncogene for CSF-1 receptor gene	AGAT
D16S539	JOE	16q24–qter	NA	GATA
D7S820	JOE	7q11.21–22	NA	GATA
D13S317	JOE	13q22–q31	NA	TATC
D5S818	JOE	5q23.3–32	NA	AGAT

[1]The August 1997 report (24,25) of the DNA Commission of the International Society for Forensic Haemogenetics (ISFH) states, "1) for STR loci within coding genes, the coding strand shall be used and the repeat sequence motif defined using the first possible 5´ nucleotide of a repeat motif; and 2) for STR loci not associated with a coding gene, the first database entry or original literature description shall be used".

[2]Amelogenin is not an STR but displays a 106-base, X-specific band and a 112-base, Y-specific band. 9947A DNA (female) displays only the 106-base, X-specific band.

TMR = carboxy-tetramethylrhodamine

FL = fluorescein

JOE = 6-carboxy-4´,5´-dichloro-2´,7´-dimethoxyfluorescein

NA = not applicable

Table 5. The PowerPlex® 16 System Allelic Ladder Information.

STR Locus	Label	Size Range of Allelic Ladder Components[1,2] (bases)	Repeat Numbers of Allelic Ladder Components	Repeat Numbers of Alleles Not Present in Allelic Ladder[3,4]
Penta E	FL	379–474	5–24	20.3
D18S51	FL	290–366	8–10, 10.2, 11–13, 13.2, 14–27	
D21S11	FL	203–259	24, 24.2, 25, 25.2, 26–28, 28.2, 29, 29.2, 30, 30.2, 31, 31.2, 32, 32.2, 33, 33.2, 34, 34.2, 35, 35.2, 36–38	
TH01	FL	156–195	4–9, 9.3, 10–11, 13.3	
D3S1358	FL	115–147	12–20	
FGA	TMR	322–444	16–18, 18.2, 19, 19.2, 20, 20.2, 21, 21.2, 22, 22.2, 23, 23.2, 24, 24.2, 25, 25.2, 26–30, 31.2, 43.2, 44.2, 45.2, 46.2	
TPOX	TMR	262–290	6–13	
D8S1179	TMR	203–247	7–18	
vWA	TMR	123–171	10–22	
Amelogenin[5]	TMR	106, 112	X, Y	
Penta D	JOE	376–449	2.2, 3.2, 5, 7–17	
CSF1PO	JOE	321–357	6–15	
D16S539	JOE	264–304	5, 8–15	
D7S820	JOE	215–247	6–14	
D13S317	JOE	176–208	7–15	
D5S818	JOE	119–155	7–16	

[1]The length of each allele in the allelic ladder has been confirmed by sequence analyses.

[2]When using an internal lane standard, such as the Internal Lane Standard 600, the calculated sizes of allelic ladder components may differ from those listed. This occurs because different sequences in allelic ladder and ILS components may cause differences in migration. The dye label also affects migration of alleles.

[3]The alleles listed are those with a frequency of >1/1000.

[4]For a current list of microvariants, see the Variant Allele Report published at the U.S. National Institute of Standards and Technology (NIST) web site at: **www.cstl.nist.gov/div831/strbase/**

[5]Amelogenin is not an STR but displays a 106-base, X-specific band and a 112-base, Y-specific band.

Table 6. The PowerPlex® 16 System Allele Determinations in Commonly Available Standard DNA Templates.

	Standard DNA Templates[1]		
STR Locus	**K562[2]**	**9947A**	**9948[3]**
Penta E	14, 5	13, 12	11, 11
D18S51	16, 15	19, 15	18, 15
D21S11	31, 30, 29	30, 30	30, 29
TH01	9.3, 9.3	9.3, 8	9.3, 6
D3S1358	16, 16	15, 14	17, 15
FGA	24, 21	24, 23	26, 24
TPOX	9, 8	8, 8	9, 8
D8S1179	12, 12	13, 13	13, 12
vWA	16, 16	18, 17	17, 17
Amelogenin	X, X	X, X	X, Y
Penta D	13, 9	12, 12	12, 8
CSF1PO	10, 9	12, 10	12, 11, 10
D16S539	12, 11	12, 11	11, 11
D7S820	11, 9	11, 10	11, 11
D13S317	8, 8	11, 11	11, 11
D5S818	12, 11	11, 11	13, 11

[1]Information on strains 9947A, 9948 and K562 is available online at: **locus.umdnj.edu/nigms/** Strain K562 is available from the American Type Culture Collection: **www.atcc.org** (Manassas, VA).

[2]Strain K562 displays three alleles at the D21S11 locus.

[3]Strain 9948 displays three alleles at the CSF1PO locus. The peak height for allele 12 is much lower than those for alleles 10 and 11.

IX.C. Power of Discrimination

The fifteen STR loci amplified with the PowerPlex® 16 System provide powerful discrimination. Population statistics for these loci and their various multiplex combinations are displayed in Tables 7–9. These data were developed as part of a collaboration (26) with The Bode Technology Group (Springfield, VA), North Carolina Bureau of Investigation (Raleigh, NC), Palm Beach County Sheriff's Office (West Palm Beach, FL), Virginia Division of Forensic Science (Richmond, VA) and Charlotte/Mecklenburg Police Department Laboratory (NC). Generation of these data included analysis of over 200 individuals from African-American, Caucasian-American and Hispanic-American populations. Data for Asian-Americans includes analysis of over 150 individuals. For additional population data for STR loci, see references 27–32.

Table 7 shows the matching probability (33) for the PowerPlex® 1.2 and 16 Systems in various populations. The matching probability of the PowerPlex® 16 System ranges from 1 in 1.83×10^{17} for Caucasian-Americans to 1 in 1.41×10^{18} for African-Americans.

A measure of discrimination often used in paternity analyses is the paternity index (PI), a means for presenting the genetic odds in favor of paternity given the genotypes for the mother, child and alleged father (34). The typical paternity indices for the PowerPlex® 1.2 and 16 Systems are shown in Table 8. The PowerPlex® 16 System provides typical paternity indices exceeding 500,000 in each population group. An alternative calculation used in paternity analyses is the power of exclusion (34). This value, calculated for the PowerPlex® 16 System, exceeds 0.999998 in all populations tested (Table 9).

Table 7. Matching Probabilities of the PowerPlex® 1.2 and 16 Systems in Various Populations.

	Matching Probability			
STR System	**African-American**	**Caucasian-American**	**Hispanic-American**	**Asian-American**
PowerPlex® 1.2 System (8 STR loci)	1 in 2.77×10^8	1 in 1.15×10^8	1 in 1.45×10^8	1 in 1.32×10^8
PowerPlex® 16 System (15 STR loci)	1 in 1.41×10^{18}	1 in 1.83×10^{17}	1 in 2.93×10^{17}	1 in 3.74×10^{17}

Table 8. Typical Paternity Indices of the PowerPlex® 1.2 and 16 Systems in Various Populations.

	Typical Paternity Index			
STR System	**African-American**	**Caucasian-American**	**Hispanic-American**	**Asian-American**
PowerPlex® 1.2 System	497	262	318	471
PowerPlex® 16 System	2,510,000	1,520,000	522,000	4,110,000

Table 9. Power of Exclusion of the PowerPlex® 1.2 and 16 Systems in Various Populations.

	Power of Exclusion			
STR System	**African-American**	**Caucasian-American**	**Hispanic-American**	**Asian-American**
PowerPlex® 1.2 System	0.9982042	0.9968863	0.9973367	0.9981793
PowerPlex® 16 System	0.9999996	0.9999994	0.9999983	0.9999998

IX.D. DNA Extraction and Quantitation Methods

The DNA IQ™ System (Cat.# DC6700) is a DNA isolation and quantitation system designed specifically for forensic and paternity samples (35). This novel system uses paramagnetic particles to prepare clean samples for STR analysis easily and efficiently and can be used to extract DNA from stains or liquid samples, such as blood or solutions. The DNA IQ™ Resin eliminates PCR inhibitors and contaminants frequently encountered in casework samples. With larger samples, the DNA IQ™ System delivers a consistent amount of total DNA. The system has been used to isolate and quantify DNA from routine sample types including buccal swabs, stains on FTA® paper and liquid blood. Additionally, DNA has been isolated from casework samples such as tissue, differentially separated sexual assault samples and stains on support materials. The DNA IQ™ System has been tested with the PowerPlex® Systems to ensure a streamlined process. See Section IX.I for ordering information.

For applications requiring human-specific DNA quantification, the AluQuant® Human DNA Quantitation System (Cat.# DC1010) has been developed (36). See Section IX.I for ordering information.

The DNA IQ™ System and AluQuant® Human DNA Quantitation System have been fully automated on the Beckman Coulter Biomek® 2000 Laboratory Automation Workstation (37). For information about automation of laboratory processes on Beckman Coulter or other workstations, contact your local Promega Branch Office or Distributor (contact information available at: **www.promega.com/worldwide/**) or e-mail: **genetic@promega.com**

IX.E. The Internal Lane Standard 600

The Internal Lane Standard (ILS) 600 contains 22 DNA fragments of 60, 80, 100, 120, 140, 160, 180, 200, 225, 250, 275, 300, 325, 350, 375, 400, 425, 450, 475, 500, 550 and 600 bases in length (Figure 12). Each fragment is labeled with carboxy-X-rhodamine (CXR) and may be detected separately (as a fourth color) in the presence of PowerPlex® 16-amplified material. The ILS 600 is designed for use in each gel lane or capillary electrophoresis (CE) injection to increase precision in analyses when using the PowerPlex® 16 System. Protocols for preparation and use of this internal lane standard are provided in Section V.

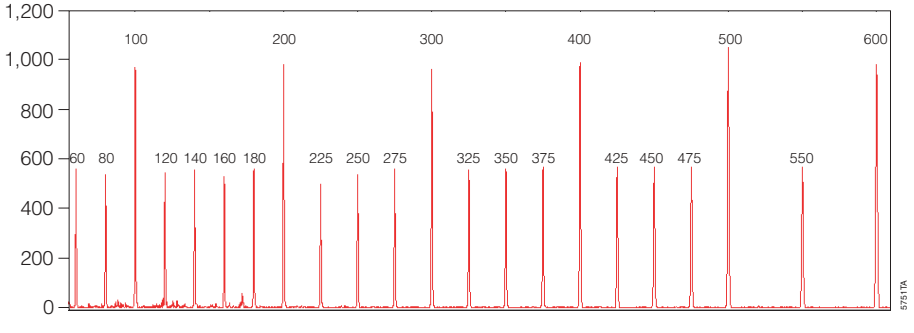

Figure 12. Internal Lane Standard 600. An electropherogram showing the fragments of the Internal Lane Standard 600.

IX.F. Preparing the PowerPlex® 16 System Master Mix

A worksheet to calculate the required amount of each component of the PCR master mix is provided in Table 10. Multiply the volume (µl) per reaction by the total number of reactions to obtain the final master mix volume (µl).

Table 10. Master Mix for the PowerPlex® 16 System.

PCR Master Mix Component	Volume Per Reaction	×	Number of Reactions	=	Final Volume (µl)
Gold ST★R 10X Buffer	2.5µl	×		=	
PowerPlex® 16 10X Primer Pair Mix	2.5µl	×		=	
AmpliTaq Gold® DNA polymerase[1]	0.8µl (4u)	×		=	
nuclease-free water[2]	µl	×		=	
Per tube		×		=	
template DNA volume[2] (0.25–1ng)	up to 19.2µl	×		=	
total reaction volume	25µl	×		=	

[1]Assumes the AmpliTaq Gold® DNA polymerase is at 5u/µl. If the enzyme concentration is different, the volume of enzyme must be adjusted accordingly.

[2]The master mix volume and template DNA volume should total 25µl. Consider the volume of template DNA, and add nuclease-free water to the master mix to bring the final volume of the final reaction to 25µl.

IX.G. Agarose Gel Electrophoresis of Amplification Products (Optional)

This procedure is optional if PCR is routinely performed in your laboratory. Agarose gel electrophoresis can be used to rapidly confirm the success of the amplification reaction prior to performing polyacrylamide gel or capillary electrophoresis.

Materials to Be Supplied by the User
(Solution compositions are provided in Section IX.H.)

- TAE 1X buffer
- agarose
- 5X loading solution
- ethidium bromide solution, 0.5µg/ml

1. Prepare a 2% agarose gel (approximately 150cm²) by adding 2.0g of agarose to 100ml of TAE 1X buffer. Mark the liquid level on the container, then boil or heat in a microwave oven to dissolve the agarose. Add preheated (60°C) deionized water to make up for any volume lost due to evaporation.
2. Cool the agarose to 55°C before pouring into the gel tray. Be sure that the gel tray is level. Pour the agarose into the tray, insert the gel comb and allow to set for 20–30 minutes.

3. Prepare the samples by mixing 10µl of each amplified sample with 2.5µl of 5X loading solution.
4. Prepare 1 liter of TAE 1X buffer for the electrophoresis running buffer.
5. Place the gel and tray in the electrophoresis gel box. Pour enough running buffer into the tank to cover the gel to a depth of at least 0.65cm. Gently remove the comb.
6. Load each sample mixed with 5X loading solution (prepared in Step 3).
7. Set the voltage at 5 volts/cm (measured as the distance between the two electrodes). Allow the gel to run for 2 hours.
8. After electrophoresis, stain the gel in TAE 1X buffer containing 0.5µg/ml ethidium bromide. Gently rock for 20 minutes at room temperature. Remove the ethidium bromide solution, and replace with deionized water. Allow the gel to destain for 20 minutes.
9. Photograph the gel using a UV transilluminator (302nm).

   Note: When analyzing the data, do not be alarmed by extra bands in addition to the alleles. DNA heteroduplexes can be expected when performing nondenaturing agarose gel electrophoresis. The sole purpose of the agarose gel is to confirm the success of the PCR reaction.

IX.H. Composition of Buffers and Solutions

10% ammonium persulfate

Add 0.05g of ammonium persulfate to 500µl of deionized water.

Blue Dextran Loading Solution

88.25%	formamide
15mg/ml	blue dextran
4.1mM	EDTA (pH 8.0)

ethidium bromide solution (10mg/ml)

1.0g	ethidium bromide

Dissolve ethidium bromide in 100ml of deionized water. Wrap in aluminum foil or transfer the solution to a dark bottle, and store at room temperature.

Caution: Ethidium bromide is a powerful mutagen. Wear gloves when working with the dye, and wear a mask when weighing it.

Gold ST★R 10X Buffer

500mM	KCl
100mM	Tris-HCl (pH 8.3 at 25°C)
15mM	$MgCl_2$
1%	Triton® X-100
2mM	each dNTP
1.6mg/ml	BSA

TAE 50X buffer (pH 7.2)

242g	Tris base
57.1ml	glacial acetic acid
100ml	0.5M EDTA stock

Add the Tris base and EDTA stock to 500ml of deionized water. Add the glacial acetic acid. Bring the volume to 1 liter with deionized water.

TBE 10X buffer

107.8g	Tris base
7.44g	EDTA ($Na_2EDTA \cdot 2H_2O$)
~55.0g	boric acid

Dissolve Tris base and EDTA in 800ml of deionized water. Slowly add the boric acid, and monitor the pH until the desired pH of 8.3 is obtained. Bring the final volume to 1 liter with deionized water.

TE^{-4} buffer (10mM Tris-HCl, 0.1mM EDTA [pH 8.0])

2.21g	Tris base
0.037g	EDTA ($Na_2EDTA \cdot 2H_2O$)

Dissolve Tris base and EDTA in 900ml of deionized water. Adjust to pH 8.0 with HCl. Bring the final volume to 1 liter with deionized water.

IX.I. Related Products

Fluorescent STR Multiplex Systems

Product	Size	Cat.#
PowerPlex® 16 Monoplex System, Penta E (Fluorescein)	100 reactions	DC6591
PowerPlex® 16 Monoplex System, Penta D (JOE)	100 reactions	DC6651
PowerPlex® ES Monoplex System, SE33 (JOE)	100 reactions	DC6751
PowerPlex® 1.2 System	100 reactions	DC6101
PowerPlex® 16 BIO System	100 reactions	DC6541
	400 reactions	DC6540
PowerPlex® ES System	100 reactions	DC6731
	400 reactions	DC6730
PowerPlex® Y System	50 reactions	DC6761
	200 reactions	DC6760

Not for Medical Diagnostic Use.

Accessory Components

Product	Size	Cat.#
PowerPlex® Matrix Standards, 310*	50µl (each dye)	DG4640
PowerPlex® Matrix Standards, 3100/3130*	25µl (each dye)	DG4650
PowerTyper™ Macros*	1 CD-ROM	DG3470
Internal Lane Standard 600**	150µl	DG2611
Gold ST★R 10X Buffer**	1.2ml	DM2411
Mineral Oil	12ml	DY1151
Nuclease-Free Water**	50ml (2 × 25ml)	P1193

*Not for Medical Diagnostic Use.
**For Laboratory Use.

Sample Preparation Systems

Product	Size	Cat.#
DNA IQ™ System**	100 reactions	DC6701
	400 reactions	DC6700
Differex™ System*	50 samples	DC6801
	200 samples	DC6800
AluQuant® Human DNA Quantitation System*	80 determinations	DC1010
	400 determinations	DC1011
Slicprep™ 96 Device**	10 pack	V1391

*Not for Medical Diagnostic Use.
**For Laboratory Use.

Polyacrylamide Gel Electrophoresis Reagents

Product	Size	Cat.#
Ammonium Persulfate	25g	V3131
TBE Buffer, 10X	1L	V4251
Urea	1kg	V3171
Blue Dextran Loading Solution*	3ml	DV4351

*For Laboratory Use.

ART® Aerosol-Resistant Tips

Product	Volume	Size (tips/pack)	Cat.#
ART® 10 Ultramicro Pipet Tip	0.5–10µl	960	DY1051
ART® 20E Ultramicro Pipet Tip	0.5–10µl	960	DY1061
ART® 20P Pipet Tip	20µl	960	DY1071
ART® GEL Gel Loading Pipet Tip	100µl	960	DY1081
ART® 100 Pipet Tip	100µl	960	DY1101
ART® 100E Pipet Tip	100µl	960	DY1111
ART® 200 Pipet Tip	200µl	960	DY1121
ART® 1000E Pipet Tip	1,000µl	800	DY1131

[a]STR loci are the subject of U.S. Pat. No. RE 37,984, German Pat. No. DE 38 34 636 C2 and other patents issued to the Max-Planck-Gesellschaft zur Förderung der Wissenschaften, e.V., Germany. The development and use of STR loci are covered by U.S. Pat. No. 5,364,759, Australian Pat. No. 670231 and other pending patents assigned to Baylor College of Medicine, Houston, Texas.

Patents for the foundational PCR process, European Pat. Nos. 201,184 and 200,362, expired on March 28, 2006. In the U.S., the patents covering the foundational PCR process expired on March 29, 2005.

[b]U.S. Pat. Nos. 6,238,863 and 6,767,703 have been issued to Promega Corporation for materials and methods for identifying and analyzing intermediate tandem repeat DNA markers. Other patents are pending.

[c]U.S. Pat. Nos. 5,843,660, 6,479,235, 6,221,598 and 7,008,771, Australian Pat. No. 724531, Canadian Pat. No. 2,118,048, Korean Pat. No. 290332, Singapore Pat. No. 57050 and Japanese Pat. No. 3602142 have been issued to Promega Corporation for multiplex amplification of STR loci. Other patents are pending.

[d]The purchase of this product does not convey a license to use AmpliTaq Gold® DNA polymerase. You should purchase AmpliTaq Gold® DNA polymerase licensed for the forensic and human identity field directly from your authorized enzyme supplier.

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